

**HEALTH-BASED MAXIMUM CONTAMINANT LEVEL  
SUPPORT DOCUMENT  
1,4-DIOXANE  
(CAS # 123-91-1; Chemical Formula: C<sub>4</sub>H<sub>8</sub>O<sub>2</sub>)**

**New Jersey Drinking Water Quality Institute  
Health Effects Subcommittee  
July 2020**

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Public Review Draft

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## ABBREVIATIONS

ALP – alkaline phosphatase  
ALT – alanine aminotransferase  
AST – aspartate aminotransferase  
BMD – Benchmark Dose  
BMR – Benchmark Response  
BW – Body weight  
CDC – Centers for Disease Control and Prevention  
CSF – Cancer Slope Factor  
DWQI – Drinking Water Quality Institute  
HED – Human Equivalent Dose  
ISGWQC – Interim specific ground water quality criterion  
ISGWQS – Interim Specific Ground Water Quality Standard  
IRIS – Integrated Risk Information System  
GWQC – Ground Water Quality Criterion  
GWQS – Ground Water Quality Standard  
LDH – lactate dehydrogenase  
LOD – Level of detection  
MAC – maximum acceptable concentration  
MCLs – Maximum Contaminant Levels  
MCLGs – Maximum Contaminant Level Goals  
MRL – Minimum Reporting Level  
NHANES – National Health and Nutrition Examination Survey  
POD – Point of Departure  
PQL - Practical Quantitation Level  
RfD – Reference dose  
SDWA – Safe Drinking Water Act  
UCMR3 – Unregulated Contaminant Monitoring Rule 3  
USEPA – United States Environmental Protection Agency

## **ABSTRACT**

This document presents the Health Effects Subcommittee's recommendation for a Health-based MCL for 1,4-dioxane. The Subcommittee's review focused primarily on the carcinogenic effects and mode of action, including the evaluation presented in USEPA IRIS (2013) and additional recent relevant information.

The Health Effects Subcommittee agreed with the USEPA Integrated Risk Information System (IRIS) (2013) conclusion that 1,4-dioxane is "likely to be carcinogenic to humans" under the USEPA (2005) Guidelines for Carcinogen Risk Assessment. The Subcommittee also agreed with USEPA IRIS (2013) that the mode of action for cancer by which 1,4-dioxane causes tumors has not been established. As specified by the USEPA (2005) guidelines, when the mode of carcinogenic action is not understood, cancer risk assessment is based on low-dose linear extrapolation (i.e. a non-threshold approach based on a cancer slope factor).

The USEPA IRIS (2013) cancer slope factor of  $0.1 \text{ (mg/kg/day)}^{-1}$  was used as the basis for the Health-based MCL. This slope factor is based on the incidence of liver tumors in female mice (Kano et al., 2009), since these tumors were the most sensitive of the tumor types caused by 1,4-dioxane in several chronic studies of male and female mice and rats.

Based on the cancer slope factor of  $0.1 \text{ (mg/kg/day)}^{-1}$ , the one in one million ( $10^{-6}$ ) cancer risk level specified in the NJ Safe Drinking Water Act, and the current USEPA default assumptions for adult body weight of 80 kg and drinking water ingestion of 2.4 L/day, a Health-based Maximum Contaminant Level (MCL) of  $0.33 \text{ }\mu\text{g/L}$  was recommended.

## EXECUTIVE SUMMARY

This document presents the Health Effects Subcommittee's recommendation for a Health-based MCL for 1,4-dioxane. To the Subcommittee's knowledge, all current U.S. federal and state ground water and drinking water guidelines for 1,4-dioxane are based on the USEPA IRIS (2013) cancer slope factor of  $0.1 \text{ (mg/kg/day)}^{-1}$ . These include the USEPA (2017) Office of Water drinking water Reference Concentration, the NJDEP (2018) Ground Water Quality Standard, and the ground water and drinking guidelines developed by 13 other states.

For this reason, the Subcommittee's review focused primarily on the carcinogenicity studies and related mode of action information for 1,4-dioxane. Other non-carcinogenic effects of this chemical were also reviewed. The Subcommittee's review used the USEPA IRIS (2013) evaluation as its starting point, and it also included more recent information identified through literature searches and submissions to the DWQI.

The Health Effects Subcommittee agreed with USEPA IRIS (2013) that, based on the occurrence of several types of tumors in multiple rodent studies, 1,4-dioxane is "likely to be carcinogenic to humans" under the USEPA (2005) Guidelines for Carcinogen Risk Assessment. According to the USEPA (2005) guidelines, risk assessment for carcinogens is based on low-dose linear extrapolation (i.e. a non-threshold approach using a cancer slope factor) when tumors occur through a mutagenic mode of action or when the mode of carcinogenic action has not been established.

The Subcommittee reviewed information relevant to 1,4-dioxane's carcinogenic mode of action including studies cited by USEPA IRIS (2013), more recent studies, and the NJDEP (2015, 2018) responses to comments on the NJDEP (2010) Interim Specific Ground Water Quality Standard and promulgated NJDEP (2018) Ground Water Quality Standard.

While many genotoxicity studies of 1,4-dioxane were negative, others provide evidence for mutagenicity and chromosomal damage. Notably, a recent *in vivo* study (Gi et al., 2018) provides evidence that 1,4-dioxane causes mutations in rat liver. Additionally, 1,4-dioxane induced micronuclei in the liver in all three studies where this effect was evaluated.

The NJDEP (2015; 2018) responses to comments includes detailed reviews of two papers (Dourson et al., 2014; Dourson et al., 2017) suggesting that 1,4-dioxane causes liver tumors through a threshold mode of action involving cell toxicity followed by regenerative growth. NJDEP (2015, 2018) concluded that the information presented in these papers does not establish a threshold mode of action for 1,4-dioxane carcinogenicity. Furthermore, the modes of action for other types of tumors (nasal, mammary gland, peritoneal) caused by 1,4-dioxane are unknown. The Subcommittee also reviewed additional information submitted by two organizations in response to the DWQI (December 2018) request for public input that questioned a non-threshold



approach for cancer risk assessment of 1,4-dioxane. The Subcommittee concluded that a mode of action for 1,4-dioxane's carcinogenicity was not established by the submitted information.

Based on its review, the Subcommittee agreed with USEPA IRIS (2013) that the mode of action for cancer by which 1,4-dioxane causes tumors has not been established. Therefore, the Subcommittee's cancer risk assessment for 1,4-dioxane was based on low-dose linear extrapolation (i.e. a non-threshold approach based on a cancer slope factor) as specified by the USEPA (2005) guidelines when the mode of carcinogenic action is not understood. The Health-based MCL is based on the USEPA IRIS (2013) cancer slope factor of  $0.1 \text{ (mg/kg/day)}^{-1}$ . This slope factor is based on the incidence of liver tumors in female mice (Kano et al., 2009), since these tumors were the most sensitive of the tumor types caused by 1,4-dioxane in several chronic studies of male and female mice and rats.

Based on the cancer slope factor of  $0.1 \text{ (mg/kg/day)}^{-1}$ , the one in one million ( $10^{-6}$ ) cancer risk level specified in the NJ Safe Drinking Water Act, and the current USEPA default exposure assumptions of adult body weight of 80 kg and drinking water ingestion of 2.4 L/day, a Health-based MCL of  $0.33 \text{ }\mu\text{g/L}$  is recommended.

## **INTRODUCTION**

### **Development of Health-based MCLs by New Jersey Drinking Water Quality Institute**

The New Jersey Drinking Water Quality Institute (DWQI) was established by the 1984 amendments to the New Jersey Safe Drinking Water Act (SDWA) at N.J.S.A. 58:12A- 20. It is charged with developing standards (Maximum Contaminant Levels; MCLs) for hazardous contaminants in drinking water and for recommending those standards to the New Jersey Department of Environmental Protection (NJDEP). The Health Effects Subcommittee of the DWQI is responsible for developing health-based drinking water levels (Health-based MCLs) as part of the development of MCL recommendations (e.g. DWQI, 1987; 1994; 2009; 2015; 2017).

Health-based MCLs are based on the goals specified in the 1984 amendments to the NJ SDWA. For carcinogens, it is generally assumed that any level of exposure results in some level of cancer risk, and a one in one million ( $10^{-6}$ ) risk level from lifetime exposure is specified in the statute. Health-based MCLs for carcinogens are thus set at levels that are not expected to result in cancer in more than one in one million persons ingesting the contaminant for a lifetime. For non-carcinogenic effects, it is generally assumed that exposure below a threshold level will not result in adverse effects as specified in the statute. Health-based MCLs for non-carcinogens are thus set at levels which are not expected to result in “any adverse physiological effects from ingestion” for a lifetime. The risk assessment approach used to develop Health-based MCLs is generally consistent with USEPA risk assessment guidance.

Other factors such as analytical quantitation limits and availability of treatment removal technology are also considered in the final MCL recommendation.

To support the development of an MCL recommendation by the DWQI, the Health Effects Subcommittee has developed a Health-based MCL for 1,4-dioxane. As specified in the 1984 Amendments to the NJ SDWA, this Health-based MCL is intended to be protective for chronic (lifetime) drinking water exposure.

### **Document Development Process**

On December 19, 2018, the DWQI announced that NJDEP Commissioner Catherine McCabe requested the DWQI to recommend an MCL for 1,4-dioxane. The Health Effects Subcommittee commenced its evaluation January 2019.

The Subcommittee began its current evaluation by reviewing the basis of the USEPA IRIS (2013) 1,4-dioxane assessment. IRIS assessments represent the scientific consensus of USEPA and undergo external peer review, and IRIS is one of the sources of toxicity factors (cancer slope factors and reference doses [RfDs]) for NJ Ground Water Quality Criteria as specified in the NJ Ground Water Quality Standard (GWQS) regulations (N.J.A.C 7:9C). IRIS evaluations have

been used as the starting point for previous Health Effects Subcommittee evaluations (for example, vinyl chloride; DWQI, 2009).

Additional information evaluated by the Health Effects Subcommittee include studies identified through literature searches as well as information submitted in response to a DWQI request for public input. At the request of the Health Effects Subcommittee, the NJDEP Environmental Library conducted three literature searches of the PubMed databases relevant studies that were not cited in the USEPA IRIS (2013) 1,4-dioxane assessment. The literature searches were performed using relevant search terms including the chemical name, CASRN and common synonyms. As discussed below, the USEPA IRIS (2013) is an update of the USEPA IRIS (2010) assessment. In the USEPA (2013) update, additional information on inhalation risk assessment was added, but the information relevant to oral exposure was not revised from USEPA IRIS (2010) which includes literature through 2009. Therefore, literature searches were performed in January 2019 for relevant citations published in 2009 through 2012 (77 citations identified) and 2013 through January 2019 (160 citations identified). An additional literature search for citations published in 2019 through March 2020 (58 citations identified) was performed in March 2020. Selection of studies for inclusion in this document were based on title and abstract screening for relevance. It is noted that the large majority of citations identified in the searches were on topics that are not relevant to the information included in this document (e.g. analytical methodologies, remediation technologies).

On December 20, 2018, the DWQI posted a request for public input for 1,4 dioxane regarding data or technical information concerning toxicology, epidemiology, toxicokinetics, or other studies related to health effects for consideration in the development of the MCL. The DWQI received three submissions, and relevant health effects comments from two of these submissions were considered by the Health Effects Subcommittee. Recent publications and information submitted in response to the DWQI request for public input questioned the use of a non-threshold (e.g. slope factor) approach for cancer risk assessment of 1,4-dioxane, based on mode of action considerations.

The Subcommittee also identified ground water and drinking water standards and guidelines for 1,4-dioxane developed by NJDEP and 13 other states. All of these standards and guidelines rely on the USEPA IRIS (2013) cancer slope factor.

As such, the Subcommittee's review focused on the carcinogenic effects of 1,4-dioxane, using the USEPA IRIS (2013) assessment as a starting point. Additional information considered included reviews of 1,4-dioxane from the peer-reviewed literature and authoritative government sources, relevant information from literature searches and screening, as well as relevant information submitted in response to the call for public input.

**BACKGROUND INFORMATION****Physical and Chemical Properties (PubChem, 2019)**

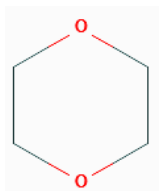
Chemical Name: 1,4-dioxane

Synonyms: diethylene ether, diethylene oxide, dioxyethylene ether, and dioxane

CAS #: 123-91-1

Chemical Formula: C<sub>4</sub>H<sub>8</sub>O<sub>2</sub>

Chemical Structure:



Molecular Weight:	88.106 g/mol
Physical State:	Liquid or solid (below 53° F)
Melting Point:	53.2 ° F/ 12° C
Boiling Point:	214 ° F/ 101° C
Vapor Pressure:	38.1 mm Hg at 25° C
Density:	1.0337 g/cm <sup>3</sup> at 20° C
Water Solubility:	>800 g/L at 25° C
Log octanol/water partition coefficient:	-0.27
Taste threshold (water):	No data
Odor threshold (water):	No data
Odor threshold (air):	170 ppm

1,4-Dioxane is a cyclic ether that exists at room temperature as a colorless liquid with a faint, pleasant ethereal odor (NTP, 2016). It is miscible with water, oils, and most organic solvents, including aromatic hydrocarbons. When it enters the air, it exists as a vapor. 1,4-Dioxane is highly flammable and may form dangerous peroxides with prolonged exposure to air and sunlight, especially in the presence of moisture (IARC, 1976; Akron, 2009).

**Production and Use**

1,4-Dioxane is a synthetic chemical used as a solvent in products such as adhesives, resins, oils and waxes; and in the pulping of wood (NTP, 2016). Historically, 90% of 1,4-dioxane was used as a stabilizer for chlorinated solvents in industrial processes, particularly 1,1,1-trichloroethane (1,1,1-TCA) (NTP, 2016; Godri Pollitt et al., 2019; ITRC 2020). The production of 1,1,1-TCA was eventually phased-out as an “ozone-depleting material” pursuant to the 1987 Montreal Protocol in the United States and 1,4-dioxane production declined (ATSDR, 2012; ITRC, 2020). Therefore, use as a solvent stabilizer for 1,1,1-TCA is no longer expected to be an important use of 1,4-dioxane (USEPA, 2013).

1,4 Dioxane is used in the manufacture of pharmaceuticals, certain plastics and rubber, and other products, and it is an impurity in antifreeze (ITRC, 2020). It is found as an unintended byproduct of surfactants used in consumer products, including personal care products and cosmetics or cosmeceuticals, and it is considered to be present as a trace contaminant in these products (ATSDR, 2012). Further uses of 1,4-dioxane include as a component of inks, paints and coatings, an additive in adhesives and a component of automotive fluids (ITRC, 2020).

Large-scale commercial production of 1,4-dioxane in the United States was first reported in 1951, but small semi-commercial quantities were available in 1929 (IARC, 1976; ATSDR, 2012). During the years 1986 and 1990, the U.S. production of 1,4-dioxane reported by manufacturers was within the range of 10–50 million pounds, and during the years 1994, 1998, and 2002, production was within the range of 1– 10 million pounds with approximately 0.9 million pounds released to the environment in 2011 (USEPA, 2011a; NTP, 2016; USEPA, 2013). More recently, 700,000 pounds of total 1,4-dioxane was released off-site in 2018 (USEPA, 2018a).

## **GUIDANCE AND STANDARDS DEVELOPED BY USEPA, NEW JERSEY AND OTHER STATES**

### **USEPA**

USEPA does not currently have an MCL for 1,4-dioxane, and it was listed on the Drinking Water Contaminant Candidate List 3 for consideration for future regulation based on its potential for public health risk and occurrence in drinking water (USEPA, 2009). No regulatory determination as to whether to pursue MCL development for 1,4-dioxane has been made by USEPA (USEPA, 2020).

The USEPA (2018b) Table of Drinking Water Health Advisories and Standards states that a concentration 35 ug/L of 1,4-dioxane in drinking water corresponds to an excess estimated lifetime cancer risk of 1 in 10,000 ( $10^{-4}$ ), based on the USEPA IRIS cancer slope factor of  $0.1 \text{ (mg/kg/day)}^{-1}$ . This slope factor is also the basis for the range of Reference Concentrations of 0.35 to 35 ug/L, based on risk levels of 1 in 10,000 ( $10^{-4}$ ) to 1 in 1 million ( $10^{-6}$ ), for evaluation of detections of 1,4-dioxane in a nationwide public water system monitoring program, the Third Unregulated Contaminant Monitoring Rule (UCMR3) (USEPA, 2017).

USEPA IRIS's initial assessment of 1,4-dioxane, posted in 1988, following the USEPA (1986) cancer risk assessment guidelines, classified the chemical as a Probable Human Carcinogen (Group B2) based on inadequate human data and sufficient evidence of carcinogenicity in animals, and an oral cancer slope factor of  $0.011 \text{ (mg/kg/day)}^{-1}$  was developed (USEPA, 1988). Following the updated USEPA (2005) risk assessment guidelines, the IRIS assessment was updated in 2010 and 1,4-dioxane was classified as "likely to be carcinogenic to humans." (USEPA, 2010). The USEPA draft 1,4-dioxane IRIS assessment was updated again to include

additional information related to the inhalation risk assessment. The risk assessment for oral exposure, including the Reference Dose and cancer slope factor, were not revised in the updated USEPA IRIS (2013) document.

### **New Jersey Health-based Drinking Water Guidance**

NJDEP Ground Water Quality Criteria (GWQC) are human health-based ground water concentrations based on drinking water exposure. As such, they are developed using the same approaches and assumptions as Maximum Contaminant Level Goals (MCLGs). The GWQC for 1,4-dioxane is based on carcinogenicity at the one-in-one million cancer risk level that is specified in the NJDEP Ground Water Quality Standard (GWQS) regulations, since carcinogenicity at this risk level is more sensitive than non-cancer effects. Also, as specified in GWQS regulations (N.J.A.C 7:9C) IRIS is one of the sources of toxicity factors that is reviewed by NJDEP in development of GWQC. NJDEP ground water quality criteria for 1,4-dioxane have been updated over time to reflect updated USEPA IRIS 1,4-dioxane assessments.

- 2008: Interim Specific Ground Water Criterion (ISGWQC) of 3 µg/L became effective in February 2008 and relied on the USEPA (1988) IRIS assessment of 1,4-dioxane.
- 2010: Revised ISGWQC of 0.35 µg/L was recommended in 2010 following NJDEP review of the USEPA IRIS (2010) updated cancer slope factor.
- 2018: NJDEP (2018) adopted a GWQS of 0.4 µg/L for 1,4-dioxane into the Ground Water Quality Standards regulations in January 2018. The earlier ISGWQS value of 0.35 µg/L was rounded to one significant figure, as specified in the NJDEP Ground Water Quality Standards regulations.

### **Other states' guidance values and standards**

Table 1 includes information on all state standards and guidance values for 1,4-dioxane in drinking water or ground water that were identified by the Health Effects Subcommittee. Of the 13 states identified, all relied on the USEPA IRIS (2013) cancer slope factor in the development of their standard or guidance value. The variations in the standards and guidance values for 1,4-dioxane are in part due to differences in the cancer risk levels (shown in Table 1) and exposure assumptions (not shown in Table 1) used by different states.

**Table 1. Levels and basis for other states' standards and guidance values for 1,4-dioxane in drinking water and ground water**

State	Standard or guidance value ( $\mu\text{g/L}$ )		Cancer slope factor	Cancer risk level
Alaska (2018)	4.6 $\mu\text{g/L}$	Groundwater cleanup	0.1 (mg/kg/day) <sup>-1</sup>	10 <sup>-5</sup>
California (2018)	1 $\mu\text{g/L}$	Notification Level (NL), health-based advisory levels	0.1 (mg/kg/day) <sup>-1</sup>	NL: 3 x 10 <sup>-6</sup> RL: 10 <sup>-4</sup>
	35 $\mu\text{g/L}$	Response Level (RL), non-regulatory		
Connecticut (2011)	3 $\mu\text{g/L}$	Action Level	0.1 (mg/kg/day) <sup>-1</sup>	10 <sup>-5</sup>
Indiana (2019)	4.6 $\mu\text{g/L}$	Groundwater screening level	0.1 (mg/kg/day) <sup>-1</sup>	10 <sup>-5</sup>
Maine (2018)	4.6 $\mu\text{g/L}$	Remedial Action Guideline	0.1 (mg/kg/day) <sup>-1</sup>	10 <sup>-5</sup>
Massachusetts (2011)	0.34 $\mu\text{g/L}$	Groundwater Standard and Non-enforceable Drinking Water Guideline	0.1 (mg/kg/day) <sup>-1</sup>	10 <sup>-6</sup>
Michigan (2017)	1.0 $\mu\text{g/L}$	Health risk limit	0.1 (mg/kg/day) <sup>-1</sup>	10 <sup>-5</sup>
Minnesota (2013)	1 $\mu\text{g/L}$	Health risk limit (Non-enforceable)	0.1 (mg/kg/day) <sup>-1</sup>	10 <sup>-5</sup>
North Carolina (2017)	0.35 $\mu\text{g/L}$	Human health criterion – Health Advisory	0.1 (mg/kg/day) <sup>-1</sup>	10 <sup>-6</sup>
New Hampshire (2018)	0.32 $\mu\text{g/L}$	Ambient Groundwater Quality Standard	0.1 (mg/kg/day) <sup>-1</sup>	10 <sup>-6</sup>
New York <sup>a</sup> (2020)	1.0 $\mu\text{g/L}$	Maximum Contaminant Level	Not applicable – see footnote below	
Texas (2009)	9.1 $\mu\text{g/L}$	Protective concentration level – cleanup standards	0.1 (mg/kg/day) <sup>-1</sup>	10 <sup>-5</sup>
Vermont (2016)	0.3 $\mu\text{g/L}$	Drinking water guidance	0.1 (mg/kg/day) <sup>-1</sup>	10 <sup>-6</sup>
Washington (2019)	0.44 $\mu\text{g/L}$	MTCA (Model Toxics Cleanup Act) Hazardous Substance	0.1 (mg/kg/day) <sup>-1</sup>	10 <sup>-6</sup>

<sup>a</sup>New York state's Drinking Water Quality Council's proposed MCL is informed by the USEPA IRIS (2013) cancer slope factor, as well as by occurrence and cost of treatment (NY DWQC, 2020).

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### **International drinking water guidelines**

Health Canada proposed a maximum acceptable concentration (MAC) of 50 µg/L for 1,4-dioxane in drinking water in August 2018 (Health Canada, 2018). The proposed MAC is based on a Tolerable Daily Intake (equivalent to a Reference Dose) for hepatic effects in rats that are stated to occur before the development of cancer. This approach is based on the assumption that there is a threshold for carcinogenicity of 1,4-dioxane, and the proposed MAC is stated to be protective of both cancer and non-cancer health effects.

The World Health Organization (WHO, 2005) also recommends a drinking water guideline of 50 µg/L. The WHO concluded that 1,4-dioxane induces multiple tumors in various organs and used a linearized multistage model for estimating cancer risk from nasal carcinomas (NCI, 1978) and hepatic tumors (Yamazaki et al., 1994) with a  $10^{-5}$  lifetime cancer risk. They also developed a second similar guideline value based on a non-cancer endpoint.

### **ENVIRONMENTAL FATE, TRANSPORT AND OCCURRENCE**

1,4-Dioxane can be released into the air, water, and soil at places where it is produced or used as a stabilizer for chlorinated solvents, particularly 1,1,1-TCA, or a solvent (Abe, 1999; ATSDR, 2012). It is expected to be degraded in the atmosphere through photooxidation with hydroxyl radical and in general is not a concern in the atmosphere since it is non-volatile and has a relatively short half-life of 35 hours (Godri Pollitt et al., 2019).

In water, 1,4-dioxane is stable and breaks down to a limited extent, if at all (ATSDR 2012; Adamson et al., 2015). 1,4-Dioxane is expected to be highly mobile in soil and is expected to leach to lower soil horizons and groundwater (ATSDR, 2012). It may be more persistent in groundwater where volatilization is hindered. 1,4-Dioxane was found in groundwater samples in the United States at concentrations ranging from 1 to 109 µg/L ppb (ATSDR, 2005). A review by Adamson et al. (2017) of 1,4-dioxane occurrence data from UCMR3, a national study of unregulated contaminants in U.S. public water systems that is discussed in detail below, concludes that 1,4-dioxane was detected almost as frequently in surface water as in ground water. However, surface water sources of 1,4-dioxane are more diluted and concentrations are generally lower than in groundwater (Adamson et al., 2017). In groundwater 1,4-dioxane is persistent with a half-life of 2-5 years, and it is less persistent in surface water with an estimated half-life of 56 days (Adamson et al., 2015; Pollitt et al., 2019).

1,4-Dioxane has been detected in contaminated surface and ground water samples collected near hazardous waste sites and industrial facilities (Derosa et al., 1996; Adamson et al., 2016). 1,4-Dioxane in ground water is highly associated with detections of chlorinated solvents, most notably 1,1,1-TCA, as well as 1,1-dichloroethane (a byproduct of 1,1,1-TCA) and trichloroethylene (TCE) (Adamson et al., 2015, 2017; Anderson et al. 2012; Godri Pollitt et al., 2019). It is suggested that the dominant source of 1,4-dioxane in the environment is from its use as a stabilizer for chlorinated solvents (Godri Pollitt et al., 2019).

1,4-Dioxane does not bioaccumulate or bioconcentrate to a significant extent in aquatic or marine organisms (ATSDR, 2012).

Based on its properties, 1,4-dioxane is not expected to partition to soil and will instead move with pore water or volatilize from dry surfaces (USEPA, 2018c).

### **Occurrence in drinking water**

Data on 1,4-dioxane in public water systems in NJ and nationwide is available through the USEPA UCMR3 (USEPA, 2017). Under UCMR3, nationwide monitoring of finished water for 30 unregulated contaminants, including 1,4 dioxane, was conducted in 2013-2015 by all U.S. large public water systems (serving more than 10,000 people) and 800 representative smaller public water systems (serving population of 10,000 or less). In UCMR3 testing, 21% of public water systems detected 1,4-dioxane (USEPA, 2017). The percentage of exceedances of the health-based Reference Level (6.9%) was higher for 1,4-dioxane than for all but one other UCMR3 contaminant (chlorate) when the Reference Level of 0.35 µg/L for 10<sup>-6</sup> cancer risk is used as the benchmark; there were no detections above the Reference Level of 35 µg/L for 10<sup>-4</sup> cancer risk (USEPA, 2017).

In UCMR3, 174 public water systems in NJ were sampled for 1,4 dioxane including 160 large systems. Of 1433 samples analyzed, 341 (23.8%) samples from 80 different public water systems were above the Minimum Reporting Level (MRL) of 0.07 µg/L. The concentrations ranged from 0.07-5.83 µg/L, with a mean concentration of 0.41 µg/L. Of the 174 water systems tested, 27 (16%) exceeded the NJ GWQS of 0.4 µg/L.

Table 2 compares UCMR3 public water system detections above the MRL and the Health Reference Concentration in New Jersey and nationally. 1,4-Dioxane was detected above the MRL of 0.07 µg/L and the Health Reference Concentration of 0.35 µg/L (which is almost identical to the NJDEP GWQS of 0.4 µg/L) more than twice as frequently in NJ than nationally.

**Table 2. New Jersey v. National Public Water System (PWS) 1,4-Dioxane Detections in UCMR3 (2013-2015)**

	New Jersey PWS		National PWS (other than NJ)	
	# Detects	% Detects	# Detects	% Detects
≥ 0.07 µg/L (MRL)	80/174	45.9%	997/4741	21.0%
≥ 0.35 µg/L (Health Reference Concentration*)	30/174	17.2%	315/4741	6.6%

\*USEPA (2017) UCMR3 Reference Concentration for 10<sup>-6</sup> cancer risk.

## **HUMAN BIOMONITORING**

The National Health and Nutrition Examination Survey (NHANES), a representative sample survey of the U.S. general population conducted by the U.S. Centers for Disease Control and Prevention (CDC, 2018), has monitored the blood concentration of 1,4-dioxane from 2009 through 2016. As 1,4-dioxane is quickly metabolized and excreted, it will not be detected unless the test is conducted within days after exposure (ATSDR, 2012; Godri Pollitt et al., 2019). In each of the four two-year cycles that have been undertaken since 2009, 1,4-dioxane in blood has been below the level of detection (LOD=0.5 ng/ml) in every participant, and more sensitive analytical methods may be required to detect the low levels of exposures in the general population (Godri Pollitt et al., 2019). Exposure to 1,4-dioxane may be evaluated through detection of its metabolites, and also tests may be available for detection of 1,4-dioxane in urine (Godri Pollitt et al., 2019).

## **SOURCES OF HUMAN EXPOSURE**

The occurrence of 1,4-dioxane in the environment primarily results from the use and disposal of associated chlorinate solvents (e.g. 1,1,1-TCA) (ITRC, 2020a).

The general public is widely exposed to 1,4-dioxane as it occurs as a byproduct in consumer products containing foaming agents, including cosmetics/toiletries, household detergents, pharmaceuticals, foods, agricultural and veterinary products, and ethylene glycol-based antifreeze coolants (Godri Pollitt et al., 2019). A 2008 survey of cosmetic products by U.S. Food and Drug Administration found 6% of product contained 1,4-dioxane between 1-5 ppm, 6% between 5-10 ppm and 8% between 10-12 ppm, while 80% of products had no detection (US FDA, 2019).

### **Drinking Water**

Drinking water is the dominant pathway of exposure to 1,4-dioxane (Godri Pollitt et al., 2019; Anderson et al., 2012). The main sources of 1,4 dioxane in drinking water are wastewater discharge, unintended spills or leaks, and historical disposal practices associated with 1,1,1-TCA (ITRC, 2020a). 1,4 -Dioxane in wastewater discharges is likely due to its widespread use in consumer products.

A two-tier multi-route exposure assessment approach concluded that exposure to 1,4- dioxane from drinking water through inhalation or dermal absorption is not significant compared to exposure from ingestion (Health Canada, 2018).

### **Food**

1,4-Dioxane has been used as a food additive and in the formulation of pesticides and food packaging adhesives (Godri Pollitt et al., 2019; ATSDR, 2012). The maximum level of 1,4-dioxane permitted in food additives (e.g. polysorbates) by the Commission of the European Communities is 5 ppm (EU Commission Directive 2003/95/EC, 2003). 1,4-Dioxane has also

been identified in several natural products including shrimp, chicken, tomatoes, coffee and certain condiments (Hartung, 1989). A study in Japan found 1,4-dioxane present in several food groups but found dietary exposure to be low (Nishimura et al., 2004).

### **Consumer Products**

Dermal exposure to 1,4-dioxane may occur through contact with residues in contaminated consumer products. 1,4-Dioxane may be a contaminant in ethoxylated surfactants, which are used in personal care products including cosmetics and shampoos and cleaning products including dishwashing liquids and detergents (Environment Canada and Health Canada, 2010). Exposure via products such as shampoo, body washes and hand soaps, which have been found to be quite high, is through inhalation and a lesser extent dermal absorption because of volatile nature of 1,4-dioxane causes most of it to evaporate (Health Canada, 2018; Environment Canada and Health Canada, 2010; ATSDR, 2012; EU, 2002). Because of their frequency of use of products containing 1,4-dioxane, women are the most exposed group (Environment Canada and Health Canada, 2010). Although low, infant exposure from consumer products has also been reported (Environment Canada and Health Canada, 2010; Godri Pollitt et al., 2019). The concentrations of 1,4- dioxane in cosmetic products have been declining over the past decade (ATSDR, 2012).

Taking effect on January 1, 2022, New York State has enacted legislation (S4389B) that will prohibit the sale of household cleaning products which contain more than trace amounts of 1,4-dioxane and will limit the sale of personal care products with certain levels of 1,4-dioxane (NYS, 2019). The legislation will phase-down permissible levels in cosmetics from 10 ppm in 1 ppm by the end of 2023.

### **Occupational**

Occupational exposure to 1,4-dioxane may occur during its production and its use as a solvent (IARC, 1999).

### **TOXICOKINETICS**

The following discussion of toxicokinetics of 1,4-dioxane is primarily based on information from USEPA (2013).

### **Absorption**

As summarized in USEPA (2013), the oral absorption of 1,4-dioxane has not been evaluated. In rats administered radiolabeled 1,4-dioxane orally, <1-2% of the radiolabel was found in the feces, indicating nearly complete oral absorption (Young et al., 1978a, b). Absorption of inhaled 1,4-dioxane in humans (Young et al., 1976, 1977) and rats (Young et al., 1978a, b) has been demonstrated by detection of the 1,4-dioxane metabolite HEAA and lower levels of unchanged 1,4-dioxane in urine after inhalation exposure. However, the fraction of 1,4-dioxane that was absorbed was not quantitated. Limited data (reviewed by USEPA, 2013) suggests that dermal absorption of 1,4-dioxane is low.

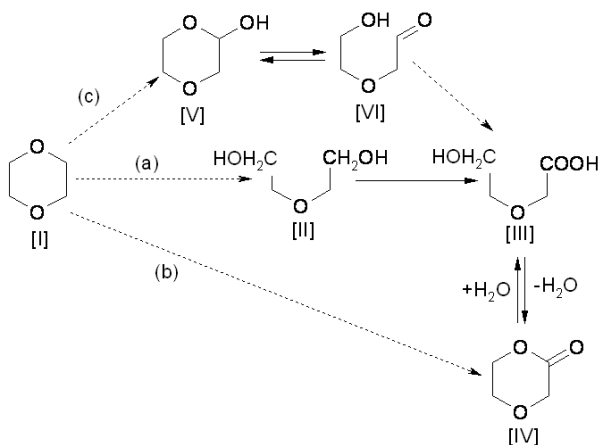
## Distribution

USEPA (2013) states that there are no available data for distribution of 1,4-dioxane in humans by any route of exposure or in animals from oral or inhalation exposure. Studies from rats injected intraperitoneally with radiolabeled 1,4-dioxane found that radiolabel was generally higher in blood than in other tissues (Woo et al., 1977; Mikheey et al., 1990).

## Metabolism

Suggested metabolic pathways of 1,4-dioxane in the rat are shown in Figure 1. Metabolism is believed to be mediated by cytochrome P-450. As summarized in USEPA (2013), the major metabolite of 1,4-dioxane in rats and humans is believed to be  $\beta$ -hydroxyethoxy acetic acid (HEAA). However, there is pH-dependent interconversion of HEAA with 1,4-dioxane-2-one, complicating interpretation of some of the studies in which these metabolites were measured (USEPA, 2013). Data from rats given a range of single gavage or intravenous doses of 1,4-dioxane indicate that metabolism is saturated as the dose increases (Young et al., 1978a, b). As discussed in USEPA (2013), the observation that metabolic saturation in rats occurred at lower doses in single-dose studies (Young et al., 1978a,b; Kociba et al., 1975) as compared to repeated-dose studies (Young et al., 1978a,b; Kasai et al., 2008) suggests that 1,4-dioxane induces its own metabolism. 1,4-Dioxane induced several isoforms of CYP450 in the liver microsomes, and one of these isoforms (CYP2E1) in nasal mucosa, and kidney microsomes, in male Sprague-Dawley rats dosed by with 2000 mg/kg/day by gavage for 2 days or 1.5% in drinking water for 10 days (Nannelli et al., 2005).

Figure 1. Suggested metabolic pathways of 1,4-dioxane in the rat.



Legend: I = 1,4-dioxane; II = diethylene glycol; III =  $\beta$ -hydroxyethoxy acetic acid (HEAA); IV = 1,4-dioxane-2-one; V = 1,4-dioxane-2-ol; VI =  $\beta$ -hydroxyethoxy acetaldehyde. Note: Metabolite [V] is a likely intermediate in pathway b as well as pathway c. The proposed pathways are based on the metabolites identified; the enzymes responsible for each reaction have not been determined. The proposed pathways do not account for metabolite degradation to the labeled carbon dioxide (CO<sub>2</sub>) identified in expired air after labeled 1,4-dioxane exposure. Source: USEPA (2013).

### **Excretion**

1,4-Dioxane and its metabolites are primarily excreted in urine in humans exposed via inhalation and in rats after inhalation or oral exposure (Young et al., 1976; 1978a,b); there are no human oral exposure data. The half-life was about one hour in both species after exposure to 50 ppm in air for 6 hours (Young et al., 1977; 1978a,b). In rats dosed orally with radiolabeled 1,4-dioxane, unchanged 1,4-dioxane and smaller amounts radiolabeled CO<sub>2</sub> (presumably 1,4-dioxane metabolites) were also found in expired air (Young et al., 1978a,b).

### **HEALTH EFFECTS – HUMAN STUDIES**

The following human health studies were identified and summarized from USEPA (2013) and EU (2002). The following evaluations, review reports or publications were also reviewed to identify additional human health studies: Health Canada (2018); ATSDR (2012); Health Council of the Netherlands (2015; carcinogenicity only) and Godri Pollitt et al. (2019). All of the studies are based on inhalation exposure; no oral studies were identified (EU 2002; Health Canada 2018).

As reviewed by USEPA (2013), EU (2002) and others, Barber (1934) was the first record of death caused by exposure to 1,4-dioxane; they reported the deaths of five patients following acute inhalation exposure to high concentrations five to eight days after symptom onset. Additionally, Johnstone (1959) further records the case of a 21-year old worker who died of kidney failure one week following inhalation and dermal exposure to high concentrations of 1,4-dioxane for one week. The liver and brain were also significantly affected as determined by autopsy (EU, 2002; USEPA, 2013).

USEPA (2013) and EU (2002) reviewed several studies of acute inhalation exposure in human volunteers. Studies reported nose and throat irritation, eye irritation and vertigo among the volunteers at varying exposure concentrations and durations (Yant et al., 1930; Silverman et al., 1946; Wirth and Klimmer 1936; Young et al., 1977). Two studies reported no symptoms after exposure (Fairley, 1934; Ernstgard et al., 2006).

Two occupational mortality studies described by USEPA (2013) found no effect from “low” exposure to dioxane (Thiess et al., 1979; Buffler et al., 1978). Thiess et al. (1979) presents a cross-sectional study which found no statistically significant effects between 74 German active and retired workers exposed to air concentrations ranging from 0.06-0.69 ppm of 1,4-dioxane. No pathological findings were reported for any of the workers. In a subset analysis of six actively employed workers and six controls there were no differences in percent of cells with gaps or other chromosome aberrations. Mortality statistics calculated for the 74 workers estimated an expected 14.5 deaths while only 12 were observed, and standardized mortality ratios for cancer did not significantly differ from the general German population. Buffler et al.,

(1978) conducted a retrospective mortality study of 165 Texas manufacturing (100) and processing (65) workers exposed for at least one month to “low” levels of 1,4-dioxane (< 25ppm). They found no statistically significant increase in cancer-related or all-cause related mortality. This study had small sample sizes among its cohorts and a short (<10-year) latency period. USEPA (2013) concluded that the two occupational 1,4- dioxane studies in humans found no conclusive causal link with increased risk for cancer (Thiess et al., 1976; Buffler et al., 1978).

Two additional studies described by EU (2002) that identified exposure but did not directly measure the concentration of 1,4-dioxane also found no effects on the health endpoints that were evaluated (Kramer et al., 1978; NIOSH, 1977). An epidemiology study of 151 employees in a textile factory, who were exposed for between one and six years to concentrations of up to 1,350 mg/m<sup>3</sup> of 1,1,1-trichloroethane blended with 4% 1,4-dioxane showed no significant differences in health including on ECG changes and liver damage, when compared to a control group (Kramer et al., 1978). Investigations on 80 men with potential exposure to 0.18 to 184 mg/m<sup>3</sup> 1,4-dioxane showed no signs of 1,4-dioxane-related health effects (NIOSH, 1977). The complete list of health endpoints evaluated by Kramer et al. (1978) and NIOSH (1977) is not provided by EU (2002).

As described in Health Canada (2018), an additional occupational study that did not directly measure 1,4-dioxane exposure reported that workers exposed to the chemicals used in silk screening and the electronics industry (known to include 1,4-dioxane) in Russia had elevated rates of spontaneous abortion and stillbirth (NIOSH, 1988; Ailamazian, 1990).

Generally noted throughout the reviews were the low quality of study reporting, in that data were obtained from secondary sources, and that study details were missing. Also, the size of the cohorts, and thus the power of the studies, was low. The potential for increased risk of cancer from occupational exposure to 1,4-dioxane could not be adequately assessed in any of the available studies.

### **HEALTH EFFECTS – ANIMAL STUDIES**

This section summarizes the toxicological information on 1,4-dioxane, including acute, short-term, subchronic and chronic oral and inhalation studies, as well as studies of reproductive/developmental and neurological effects. In most of the repeated-dose studies, 1,4 dioxane caused toxicity to the liver, kidney and respiratory tract. It caused liver tumors in multiple studies in rats, mice and guinea pigs, as well as nasal tumors in rats exposed through drinking water or inhalation. Increases in incidence of several other types of tumors were also reported in one or more rat studies. It should be noted that studies that evaluated the carcinogenicity of 1,4-dioxane in laboratory animals, regardless of duration (e.g. subchronic or chronic) are discussed in the section on “Chronic studies and other studies evaluating

carcinogenicity” below. The summaries of these studies also include the non-neoplastic effects that were reported.

### **Acute and short-term studies**

LD<sub>50</sub> values from a single gavage dose of 1,4-dioxane have been reported as follows: rat: 5,400-7,210 mg/kg (Laug et al., 1939; Pozzani et al., 1959; Smyth et al., 1941); mouse: 5,900 mg/kg (Laug et al., 1939); guinea pig: 3,150-4,030 mg/kg (Laug et al., 1939; Smyth et al., 1941).

Toxicological effects observed in acute (single dose) and short-term (up to 14 day) gavage and drinking water studies are summarized in USEPA (2013). Histopathological changes in the liver and kidney were reported in rats (David, 1964; Kesten et al., 1939, JBRC, 1998; Kitchin and Brown, 1990; Nelson, 1951), mice (Laug et al., 1939; JBRC, 1998), guinea pigs (Laug et al., 1939), rabbits (de Navasquez, 1935), and dogs (Schrenk and Yant, 1936). Nasal and brain lesions in rats and mice were also reported in the 14-day drinking water study conducted by the Japan Bioassay Research Center (JBRC, 1998).

### **Subchronic studies**

#### Oral studies

As summarized by USEPA (2013), Fairley et al. (1934) dosed 6 rats and 6 mice with drinking water containing 1.25% (12,500 ppm) 1,4-dioxane for up to 67 days, resulting in estimated doses of 1,900 mg/kg/day in rats and 3,300 mg/kg/day in mice. Only one rat survived past day 34, while 5 mice survived until day 60. Effects associated with treatment in both species included enlarged kidneys and histopathological changes in the kidney and liver.

As summarized by USEPA (2013), Stott et al. (1981) dosed male Sprague-Dawley rats (4-6 per group) with 0, 10, or 1,000 mg/kg/day in their drinking water, 7 days/week for 11 weeks. It was noted by USEPA (2013) that the high dose was stated to be 100 mg/kg-day in the Methods section, while the Abstract, Results, and Discussion sections state that it was 1,000 mg/kg-day. As such, it was assumed to be 1000 mg/kg/day. In the high-dose group, relative liver weight was increased, histopathological changes were observed in the liver, and hepatic DNA synthesis, measured by [<sup>3</sup>H]-thymidine incorporation, was increased 1.5-fold. No effects were reported in the low-dose group.

Kano et al. (2008) dosed F344/DuCrj rats (10/sex/group) and Crj:BDF1 mice (10/sex/group) with 0, 640, 1,600, 4,000, 10,000, or 25,000 ppm 1,4-dioxane in drinking water for 13 weeks. In rats, daily doses were estimated based on water consumption and body weight data as 0, 52, 126, 274, 657, and 1,554 mg/kg/day in males, and 0, 83, 185, 427, 756, and 1,614 mg/kg/day in females. One female in the high dose group died; the cause and time of death were not provided. Body weights at the end of the study were decreased at the two highest dose levels in females (12 and 21%) and males (7 and 21%), respectively. Food consumption was reduced in the highest



dose group by 13% in females and 8% in males, and water consumption was decreased in a dose-related fashion at all doses in males and starting at the second lowest dose level in females. Red blood cells, hemoglobin and hematocrit were significantly increased in high dose males but were not affected in females. The liver enzymes AST and ALT were significantly increased in high dose males, and AST was also increased in high dose females, and plasma glucose was decreased in high dose males and females. Absolute and relative kidney weights were increased in females in all but the lowest dose group. Histopathological changes were reported in respiratory, olfactory and tracheal epithelium, liver, kidneys and brain. The most sensitive effects were nuclear enlargement of the nasal cavity respiratory epithelium and hepatocyte swelling, occurring at 126 mg/kg/day in males.

In mice, daily doses were estimated based on water consumption and body weight data as 0, 86, 231, 585, 882, or 1,570 mg/kg/day in males, and 0, 170, 387, 898, 1,620, or 2,669 mg/kg/day in females. One male mouse in the high-dose group died; the cause and time of death were not provided. Body weights at the end of the study were decreased by 29% in high dose males, and by less than 10% in other dosed groups; food consumption was not affected. Water consumption was decreased in all dosed groups of males and in the highest dose group of females, with 70% and 57% decreases at the highest dose in males and females, respectively. As was the case in rats, red blood cells, hemoglobin and hematocrit were significantly increased in high dose males but were not affected in females. The liver enzymes AST and ALT were significantly increased in high dose males and females. Plasma glucose was decreased in high dose males and at the two highest doses in females. Absolute and relative lung weights were increased in high dose males and in the two highest dosed groups of females. Absolute kidney weight was increased in the two highest dosed groups of females, and relative kidney weight was also increased in the highest female dosed group. Histopathological changes were reported in respiratory, olfactory, and hepatic tissues. The most sensitive endpoint reported by the authors was nuclear enlargement in the bronchial epithelium in females at 387 mg/kg/day; USEPA (2013) notes that it does not consider nuclear enlargement to be an adverse effect.

An additional recent subchronic study focused on renal effects of 1,4-dioxane. Qiu et al. (2019) administered 0, 0.5 or 500 ppm 1,4-dioxane in drinking water to groups of 12 male mice for 12 weeks. Estimated doses were 0.1 and 100 mg/kg/day. While food and water consumption were not affected by treatment, body weight gain was decreased at both doses and relative kidney weight was increased at the higher dose. No histopathological changes were reported in the kidney at the low dose, while hydropic generation of the renal tubules, glomerular cell proliferation, hyperemia and slight inflammation were observed at the high dose. Other components of the study evaluated indicators of oxidative stress (superoxide dismutase and glutathione) in renal tissue, urinary protein and creatinine, transcriptomic analysis of renal tissue and metabolomic analysis of urine. Although there were no histopathological effects or changes in indicators of oxidative stress in the low dose group, transcriptomic changes in signaling pathways related to oxidative stress and other biological processes occurred, suggesting that

oxidative damage may be occurring even in the absence of more overt effects. Metabolomic and transcriptomic data also indicated effects on metabolic pathways, including those involving amino acid metabolism.

### Inhalation studies

As summarized by USEPA (2013), Fairley et al. (1934) exposed rats, mice, guinea pigs and/or rabbits (3–6/species/group) to 1,000, 2,000, 5,000, or 10,000 ppm of 1,4-dioxane vapor for varying lengths of time, twice a day for 1.5 hours for 5 days/week and once for 1.5 hours on the sixth day of the week. At 10,000 ppm, all animals except one rat died within the first five exposures. At 5,000 ppm, two mice and one guinea pig died after 15–34 exposures while the remaining animals were sacrificed after 3 weeks or 5 weeks of exposure. At 2000 and 1000 ppm, animals were exposed for approximately 2–6 weeks and 4–12 weeks, respectively. Kidney and liver damage occurred at all doses and was more severe at higher doses.

As summarized by USEPA (2013), Kasai et al. (2008) exposed F344/DuCrj rats (10/sex/group) to 0, 100, 200, 400, 800, 1,600, 3,200, or 6,400 ppm of 1,4-dioxane for 6 hours/day, 5 days/week, for 13 weeks. At the highest dose, all rats died by the end of the first week from renal failure caused by necrosis of the renal tubules; no deaths occurred at lower concentrations. Statistically significant increases in several organ weights included lungs ( $\geq 1,600$  ppm, males;  $\geq 200$  ppm, females); livers ( $\geq 800$  ppm, both sexes), and kidneys (3,200 ppm, males;  $\geq 800$  ppm, females). Statistically significant changes in hematological parameters and clinical chemistry at 3,200 ppm included increased hemoglobin ALT, erythrocytes, AST, and mean corpuscular volume in both sexes; increased hematocrit in females; and decreased glucose and triglyceride in males were observed. Histopathological changes caused by 1,4-dioxane in one of both sexes were reported in the nasal respiratory, nasal olfactory, tracheal, and bronchial epithelium; kidney; and liver. Glutathione S-transferase placental form (GST-P) foci, a preneoplastic liver lesion, was found in 3/10 males and 2/10 females at 3,200 ppm and 4/10 females at 1,600 ppm. The authors identified nuclear enlargement in the respiratory epithelium of males and females at 100 ppm as the most sensitive endpoint. As noted above, USEPA (2013) stated that they do not consider this to be an adverse effect.

### Studies of neurological effects

Clinical signs of CNS depression (e.g. staggered gait, narcosis, paralysis, coma, and death) were observed in some of the studies mentioned above. USEPA (2013) reviewed four rodent studies that focused on specific neurological effects of 1,4-dioxane, including one oral study (Goldberg et al., 1964) and two inhalation studies (Frantik et al., 1994; Kanada et al., 1994).

### Oral studies

In male rats administered a single gavage dose of 1,050 mg/kg, dopamine and serotonin levels were reduced compared to controls in the hypothalamus, while no effects were seen in other parts of the brain (Kanada et al., 1994).

### Inhalation studies

Frantik et al. (1994) evaluated the effect of inhaled 1,4-dioxane on the propagation and maintenance of an electrically-evoked seizure discharge in rats and mice. The most sensitive and reproducible effects were duration of tonic hind limb extension in rats and the velocity of tonic extension in mice. The 1,4-dioxane air concentration producing a 30% decrease in the maximal response to an electrically-evoked seizure was  $1,860 \pm 200$  ppm in rats and  $2,400 \pm 420$  ppm in mice, and no NOAEL was identified.

Goldberg et al. (1964) evaluated the effect of 1,4-dioxane inhalation on conditioned avoidance and escape behaviors using a pole climb methodology. A dose-related effect on conditioned avoidance behavior was observed in female rats exposed to 0, 1,500, 3,000, or 6,000 ppm of 1,4-dioxane in air for 10 days, 4 hours/day, 5 days/week. In the high dose group where the effect was greatest, a higher percentage of rats were affected during the first 2 days of exposure than on days 3-10.

### Study of reproductive and developmental effects

Only one study of reproductive and developmental effects of 1,4-dioxane was identified. As summarized by USEPA (2013), Giavini et al. (1985) administered 0, 250, 500, or 1000 mg/kg/day 1,4-dioxane by gavage to pregnant female Sprague Dawley rats (18–20 per dose group) on gestations days 6–15 and sacrificed them on gestation day 21. In the high dose group, fetal weight was decreased by 5% ( $p < 0.01$ ) and ossification of the sternebrae was reduced ( $p < 0.05$ ), while maternal weight gain was decreased by 10% in this dose group. Numbers of corpora lutea, implantations, resorptions, and live fetuses and external, visceral, and skeletal fetal malformations were not affected by treatment. The study authors suggested that delayed ossification and decreased fetal weight indicate a developmental delay at 1000 mg/kg/day.

### Chronic studies and other studies evaluating carcinogenicity

As shown in Table 3, 1,4-dioxane caused tumors in multiple organs in studies of rats, mice and guinea pigs. These studies are summarized below.

Table 3. Sites at which tumor incidence was increased by 1,4-dioxane in animal studies (adapted from USEPA, 2013)

Study Details				Tumor Sites								
Study	Exposure Route	Species	Sex	Liver	Nasal Cavity	Mammary Gland	Peritoneal Mesothelioma	Testis/Epididymis Mesothelioma	Kidney	Zymbal Gland	Subcutis Fibroma	
<b>Argus et al. (1965)</b>												
	Drinking Water	Rat	M	+	-	-	-	-	-	-	-	
<b>Hoch-Ligeti et al. (1969) / Argus et al. (1973)</b>												
	Drinking Water	Rat	M	+	+	NR*	NR	NR	NR	NR	-	
<b>Hoch-Ligeti and Argus (1970)</b>												
	Drinking Water	Guinea Pig	M	+	-	-	-	-	-	-	-	
<b>Kociba et al. (1974)</b>												
	Drinking Water	Rat	M/F	+	+	-**	-	-	-	-	-	
<b>NCI (1978)</b>												
	Drinking Water	Rat	M	-	+	-	-	+***	-	-	-	
			F	+	+	-	+	NA	-	-	-	
		Mouse	M	+	-	-	-	-	-	-	-	-
			F	+	-	-	-	-	NA	-	-	-
<b>Kano et al. (2009)</b>												
	Drinking Water	Rat	M	+	+	+	-	-	-	-	-	
			F	+	+	+	+	NA	-	-	-	
		Mouse	M	+	-	-	-	-	-	-	-	-
			F	+	-	-	-	-	NA	-	-	-
<b>Torkelson et al. (1974)</b>												
	Inhalation	Rat	M	-	NR	-	-	-	-	-	-	
			F	-		-	-	-	-	-		
<b>Kasai et al. (2009)</b>												
	Inhalation	Rat	M	+	+	+	-	-	+	+	+	
*NR –not reported. **Some organs marked “-” may not have been evaluated. ***Increase was noted as not statistically significant. not reported.												

### Oral studies

Argus et al. (1965) administered drinking water containing 1% (10,000 ppm) 1,4-dioxane to 26 adult male Wistar rats for 63 weeks, resulting in a dose estimated by USEPA (2013) of 640 mg/kg/day. There were 9 rats in the control group. Liver tumors occurred in 6/26 treated rats, while there were no liver tumors in the control group. Histopathological changes in the liver were observed in rats that died before the end of the dosing period, beginning at 2.5 weeks after dosing began. Extensive histopathological changes in the kidney also occurred in “many” treated rats (incidence not provided).

Stoner et al. (1986) included 1,4-dioxane in a study of 19 chemicals that compared the induction of lung tumors in A/J mice from exposure via gavage versus intraperitoneal injection. Groups of 16 male and female mice were dosed 3 times per week for 8 weeks with average daily doses that were estimated by USEPA (2013) as 430 mg/kg/day for gavage dosing and 86, 210, or 430 mg/kg/day for intraperitoneal dosing. The mice were sacrificed 24 weeks after dosing began. It was concluded that 1,4-dioxane did not induce lung tumors by either route of exposure in this subchronic study.

Hoch-Ligeti et al. (1969) and Argus et al. (1973) reported on a study in which groups of 28-32 male Charles River CD rats (two to three months old) were administered drinking water containing 0, 0.75, 1.00, 1.40 or 1.80 % of 1,4-dioxane to for 13 months. Doses were estimated by USEPA (2013) as 0, 430, 574, 803, and 1032 mg/kg/day. Animals were sacrificed at 16 months, or earlier if nasal cavity tumors were observed. An additional group of 10 rats was exposed to 1% 1,4-dioxane for electron microscopy studies of the liver after exposure for 5 months (5 rats) or 13 months (5 rats). Tumor data were reported only for the nasal cavity (Hoch-Ligeti et al., 1969) and the liver (Argus et al., 1973). Only nasal tumors visible from gross examination were reported, and histological examination of the nasal cavity was not performed on rats without visible nasal tumors. The number of nasal cavity tumors per group (28-32 rats) was: 0 mg/kg/day – 0; 430 mg/kg/day – 1; 574 mg/kg/day – 1; 803 mg/kg/day – 2; 1032 mg/kg/day – 2. These tumors were observed between approximately 11-16 months after dosing began. In the liver, the incidence of “incipient tumors” (nodules showing all of the histological characteristics of fully developed hepatomas) and hepatomas was reported in the treated groups, but liver tumor data were not provided for the control group. The number of liver tumors increased with dose as follows: 430 mg/kg/day – 4 incipient tumors, 0 hepatomas, 4 total tumors; 574 mg/kg/day – 9 incipient tumors, 0 hepatomas, 9 total tumors; 803 mg/kg/day – 13 incipient tumors, 3 hepatomas, 16 total tumors; 1032 mg/kg/day – 11 incipient tumors, 12 hepatomas, 23 total tumors. Two other types of nodules (one consisting of large cells with reduced cytoplasmic basophilia, the other consisting of large cells filled with fat) often occurred in the livers that had tumors. The electron microscopy studies showed changes in the liver cell ultrastructure comparable to those caused by other hepatic carcinogens (aflatoxin B1; dialkyl nitrosamines, and others), with effects progressing between 8 and 13 months of exposure. In addition to the nasal cavity and liver tumors, all dose levels of 1,4-dioxane caused notable histopathological changes in the kidney (incidence not reported).

Hoch-Ligeti and Argus (1970), as summarized by USEPA (2013), provide a “brief account” of a study in which 22 male guinea pigs were exposed to 1,4-dioxane in drinking water for 23-28 months at concentrations varying from 0.5 – 2% over time. USEPA (2013) estimated the dose to 944 – 1019 mg/kg/day. The control group consisted of 10 guinea pigs. In the treated group, two guinea pigs had carcinoma of the gallbladder, three had early hepatomas, and one had a renal

adenoma. The incidence of histopathological changes in the lungs was increased in the dosed group.

Kociba et al. (1974) administered 0, 0.01, 0.1 or 1.0% 1,4-dioxane in drinking water to groups of 60 male and female 6 to 8-week-old Sherman rats for up to 716 days (102 weeks). Based on measured water consumption and body weight data, mean daily doses were calculated as 0, 9.6, 94 and 1015 mg/kg/day in males and 0, 19, 148 and 1599 mg/kg/day in females. Body weight was decreased in the high dose group throughout the study. Mortality in high dose males and females was increase significantly in the first 4 months of the study, with less than 60% of rats surviving, and the rats that died showed degenerative changes in the liver and kidney. The rate of mortality did not differ substantially between control and treated groups starting at month 5, but due to the high early mortality, only 1 male and a small number of females survived until the end of the study. There were no effects on hematological parameters evaluated at 4, 6, 12, 18, and 24 months. The only significant organ weight change was increased absolute and relative liver weight in the few high dose rats that survived until the end of the study. Non-neoplastic histopathological changes were reported in the kidneys (renal tubular epithelial degeneration and regenerative activity) and livers (hepatocellular degeneration and necrosis; hyperplastic nodules) in the mid- and high-dose groups, but not in the low dose groups (incidence not reported). Hepatocellular carcinomas occurred in 1 control, no low dose, 1 mid-dose, and 10 (6 male, 4 female) high dose rats, and nasal carcinomas occurred only in 3 (1 male, 2 female) high-dose rats. The statistical analysis of tumor incidence presented by the authors is based on the number of rats surviving at 12 months, since almost all of the tumors (including all hepatic tumors) were noted at 12 months or later. Total hepatic tumors ( $p=0.00022$ ), hepatocellular carcinomas ( $p=0.00033$ ), and nasal carcinomas ( $p=0.05491$ ) were significantly increased in high-dose rats (males and females combined).

The National Cancer Institute (NCI, 1978) conducted a chronic study in which 0, 0.5 or 1% 1,4-dioxane in drinking water was administered to groups of 35 male and female Osborne-Mendel rats were dosed for 110 weeks and groups of 50 male and female mice B6C3F1 mice for 90 weeks (approximately 4 weeks old). Dosing of the control and high-dose male rats started 1 year after the study began, due to death of the original groups due to an air-conditioning failure. Therefore, the study of the control and high dose males took place at a different time than the study of the females and low dose males.

In rats, mean daily doses were calculated as 0, 240 and 530 mg/kg/day in males, and 0, 350 and 640 mg/kg/day in females, based on measured water consumption and body weight data. There was a statistically significant dose-related increase in mortality in both males and females. Non-neoplastic lesions that were significantly increased in treated groups included renal cortical tubular degeneration (low- and high-dose males; high-dose females), hepatocytomegaly (high-dose females), gastric ulcers (low- and high-dose males), and pneumonia (high-dose females). In treated rats, there was an increased incidence of tumors of the nasal cavity (squamous cell

carcinomas, adenocarcinomas, and one rhabdomyoma) in males and females, liver (hepatocellular adenomas) in females, and testis/epididymis (mesotheliomas; not statistically significant) in males. The first tumors were observed at week 52 in males and week 66 in females. Of these tumor types, the increases in nasal cavity squamous cell carcinomas (0/33, 12/33, 16/34 in control, low-dose, and high-dose females; 0/34, 10/35, 8/35 in control, low-dose, and high-dose males) were statistically significant, as was the increase in hepatocellular adenomas in females (0/31, 10/33, 11/32 in control, low-dose, and high-dose).

In mice, mean daily doses were calculated as 0, 720 and 830 mg/kg/day in males, and 0, 380 and 860 mg/kg/day in females, based on measured water consumption and body weight data. There was a statistically significant dose-related increase in mortality in females beginning at about week 80, while mortality was not affected by treatment in males. The authors stated that differences in body weight between controls and dosed groups in the second year may have been due to fluctuations within the smaller numbers of remaining mice surviving until this time. Non-neoplastic lesions that were significantly increased in treated groups included pneumonia in males and females, and rhinitis in females. In a statistical analysis performed by USEPA (2013), the incidence of pneumonia and rhinitis in low-dose and high-dose females compared to controls was significantly increased at  $p < 0.001$ . In treated mice, the incidence of hepatocellular carcinomas and hepatocellular adenomas or carcinomas was significantly increased in a dose-related manner in all treated groups. In males, the incidence of carcinomas was 2/49, 18/50, and 24/27, and the incidence of adenomas or carcinomas was 8/49, 19/50, and 28/47, in the control, low-, and high-dose groups, respectively. In females, the incidence of carcinomas was 0/50, 12/48, and 29/37, and the incidence of adenomas or carcinomas was 0/50, 21/48, and 35/37, in the control, low-dose, and high-dose groups, respectively.

Kano et al. (2009) reported on a study in which groups of 50 male and female F344/DuCrj rats and groups of 50 male and female Crj:BDF1 mice were exposed to drinking water containing 0, 500, 2000 or 8000 ppm 1,4-dioxane for 2 years. This study was also reported in a Japan Bioassay Research Center report (JBRC, 1998) and in conference proceedings by Yamazaki et al. (1994).

In rats, mean daily doses were calculated as 0, 11, 55, and 274 mg/kg/day in males, and 0, 18, 83, and 429 mg/kg/day in females, based on measured water consumption and body weight data. Growth rates and terminal body weights were significantly lower in male and female high-dose rats than controls, although food consumption was not affected by treatment. No mortality occurred in controls or treated rats during the first 12 months of the study. At the end of the two-year study, only about 50% of high-dose males and females survived, and survival in this group was significantly lower than in the controls. Kano et al. (2009) attributed the lower survival in the high-dose groups to deaths due to nasal tumors and peritoneal mesotheliomas in males, and nasal and hepatic tumors in females.

USEPA (2013) summarized the hematology and clinical chemistry parameters that were evaluated at the end of the two-year study and reported by JRBC (1998). Decreases in red blood cells, mean corpuscular volume, hemoglobin and hematocrit, and increases in platelets, occurred in high-dose males and females; all of these effects except increased mean corpuscular volume also occurred in mid-dose males. There were significant changes in serum chemistry parameters in the high dose groups. In males, these included increased phospholipids, AST, ALT, LDH, ALP, GGT, CPK, potassium and inorganic phosphorus, and decreased total protein, albumin, and glucose. In females, changes included increased total bilirubin, cholesterol, phospholipids, AST, ALT, LDH, GGT, ALP, CPK and potassium, and decreased blood glucose. Serum enzyme activity was increased by <2 to 17-fold compared to controls, with the largest increases for ALT, AST and GGT. Relative liver weight was increased in mid-dose males and in high-dose males and females.

Data on non-neoplastic histopathological lesions presented by JBRC (1998) and Kano et al. (2009) are summarized in USEPA (2013). Effects occurred in the nasal cavity, liver and kidneys, primarily in the high-dose groups with some effects also seen in the mid-dose groups. Nasal cavity lesions in high-dose males included nuclear enlargement and metaplasia of the olfactory and respiratory epithelia, atrophy of the olfactory epithelium, hydropic changes and sclerosis of the lamina propria, adhesion, and inflammation. In female rats, nuclear enlargement and metaplasia of the respiratory epithelium, squamous cell hyperplasia, respiratory metaplasia of the olfactory epithelium, hydropic changes and sclerosis of the lamina propria, adhesion, inflammation, and proliferation of the nasal gland occurred in the high-dose group, and nuclear enlargement of the olfactory epithelium occurred in both the mid- and high-dose groups. In the liver, spongiosis hepatitis and clear and mixed cell foci occurred in mid- and high-dose males, and spongiosis hepatitis, cyst formation and mixed cell foci occurred in high-dose females. In the kidney, nuclear enlargement of the renal proximal tubule occurred in high-dose males and mid- and high-dose females.

Tumors of the liver, nasal cavity and mammary gland were significantly increased in high-dose males and females, and peritoneal mesotheliomas were also increased in high-dose males. Liver tumors were observed beginning at earlier time points in high-dose males and females than in lower dose groups and controls. In high-dose males, the incidence of hepatocellular adenomas and either adenoma or carcinoma were 32/50 and 39/50, respectively, compared to 3/50 for both parameters in controls. Nasal cavity squamous cell carcinomas occurred in 3/50 and 7/50 high-dose males and females, while the incidence in male and female controls was 0/50. Peritoneal mesotheliomas occurred in 28/50 high-dose males and 2/50 controls, and these tumors were the most frequent cause of death in high-dose males. In high dose males, mammary gland fibroadenomas (4/50) and either fibroadenoma or adenoma (6/50) were increased compared to controls (1/50 for both parameters). Similarly, in high-dose females, mammary gland adenomas (16/50) and adenomas or fibroadenomas (18/50) were increased compared to controls (3/50 and 8/50, respectively).



In mice, mean daily doses were calculated as 0, 49, 191, and 677 mg/kg/day in males, and 0, 66, 278, and 964 mg/kg/day in females, based on measured water consumption and body weight data. Growth rates and terminal body weights were significantly lower than control in mid-dose and high-dose males and females, with decreases of 43% and 45%, respectively, in the high-dose males and females. However, food consumption was not significantly affected by treatment. Survival was not affected by treatment in males, but survival in mid- and high-dose females was significantly lower than in controls. Almost all of the deaths in the treated groups occurred during the second year of the study, and most of these deaths were attributed to hepatic tumors.

USEPA (2013) summarized the hematology and clinical chemistry parameters that were evaluated at the end of the two-year study and reported by JRBC (1998). Red blood cell numbers, hemoglobin, and hematocrit were increased in males, and platelets were decreased in mid- and high-dose males and females. Clinical chemistry changes included, among others, increased AST, ALT, LDH, and ALP activities in mid- and high-dose females, and increased CPK activity in high-dose females.

Absolute and relative lung weights were increased in high-dose males and in mid- and high-dose females (JBRC, 1998, reported in USEPA, 2013). Non-neoplastic histopathological changes were observed in the epithelium of the respiratory tract in high-dose and some mid-dose mice and in the proximal tubule of the kidney in high-dose males, as well as and angiectasis (dilation of blood vessels) in the liver in high-dose males.

An increased incidence of liver tumors (adenomas and carcinomas) occurred in treated male and female mice. In males, the incidence of hepatocellular carcinoma and adenoma or carcinoma) was significantly increased in all dose groups, and the incidence of adenoma was statistically increased only in the mid-dose group only. In female mice, the incidence of hepatocellular carcinoma was significantly increased in all dosed groups, and hepatocellular adenoma was increased in the low- and mid-dose groups.

#### Inhalation studies

Torkelson et al. (1974) exposed male and female Wistar rats (288 per sex) to 111 ppm 1,4-dioxane in whole body inhalation chambers for 7 hours/day, 5 day/week for 2 years. There were 192 controls per sex. 1,4-Dioxane exposure did not affect mortality, body weight gain or organ weights. Slight, but statistically significant, changes in hematology and clinical chemistry parameters were within normal limits and were not considered to be toxicologically relevant by the investigators. No non-neoplastic histopathological changes were associated with treatment. The incidence of various types of tumors did not differ significantly between the control and treated groups, although it is noted that nasal cavity tumors were not evaluated.

Kasai et al. (2009) exposed 6-week-old male F344/DuCrj rats (50 per group) to 0, 50, 250 or 1250 ppm 1,4-dioxane in whole body inhalation chambers for 6 hours/day, 5 day/week for 2 years. 1,4-Dioxane did not significantly affect growth rates during the first 5 months of the study, but growth was decreased in all treated groups during the second year of the study, while food consumption was not affected. Survival was significantly decreased following 91 weeks of exposure to 1,250 ppm of 1,4-dioxane, and these deaths were attributed primarily to increased incidences of peritoneal mesotheliomas, with nasal tumors also contributing. Statistically significant changes in hematology and clinical chemistry parameters in the high-dose group at the end of the two-year study included decreased hemoglobin, mean corpuscular volume and mean corpuscular hemoglobin, and increased AST, ALT, ALP and  $\gamma$ -GTP.

Non-tumor histopathological changes occurred at an increased incidence in all dose groups in the nasal cavity, in the mid- and high-dose groups in the kidney, and in the high-dose group in the liver. There was a significant increase in multiple types of tumors. These included squamous cell carcinomas of the nasal cavity, hepatocellular adenomas, renal cell carcinomas, mammary gland fibroadenomas, peritoneal mesotheliomas, and Zymbal gland adenomas. The tumor types with the highest incidence in the high-dose group were peritoneal mesotheliomas (41/50 compared to 2/50 in controls) and hepatocellular adenomas (21/50 compared to 1/50 in controls).

## **MODE OF ACTION**

### **Genotoxicity**

The genotoxicity of 1,4-dioxane has been evaluated in numerous *in vitro* and *in vivo* studies. USEPA (2013) provides a detailed review of the genotoxicity data for 1,4-dioxane that were available at the time that it was written. Two more recent *in vivo* studies, Gi et al. (2018) and Itoh and Hattori (2019), were identified by the Health Effects Subcommittee.

As summarized by USEPA (2013), the majority of *in vitro* genotoxicity tests gave negative results. A number of bacterial mutagenicity assays, with and without metabolic activation, were negative. However, it should be noted that a recent *in vivo* study (Gi et al., 2018 – discussed below) provides evidence that 1,4-dioxane causes mutations in rat liver. Other negative genotoxicity studies include an evaluation of induction of aneuploidy in yeast, a sex-linked recessive lethal test in *Drosophila melanogaster* (fruit flies), a mouse lymphoma forward mutation assay, a study of chromosomal aberrations and micronucleus formation in Chinese hamster ovary (CHO) cells, a study of sister chromatic exchange in CHO cells, and a study of *in vitro* covalent binding to DNA.

Positive *in vitro* genotoxicity studies reported increased meiotic non-disjunction in *Drosophila* oocytes (Munoz and Barnett, 2002), increased single strand DNA breaks in rat hepatocytes at concentrations that decreased cell viability (Sina et al., 1983), increased sister chromatid exchange in CHO cells only at the highest dose tested without metabolic activation and not at

any dose with metabolic activation (Galloway et al., 1987), and increased transformation of BALB/3T3 cells accompanied by toxicity (Sheu et al, 1988).

Data on micronucleus formation in peripheral blood and bone marrow are mixed. 1,4-Dioxane did not cause micronucleus formation in the bone marrow of B6C3F1, BALB/c, CBA, or C57BL6 mice (McFee et al., 1994; Mirkova, 1994; Tinwell and Ashby, 1994) and male F344/DuCrIj rats (Itoh and Hattori, 2019), or in peripheral blood of CD1 mice (Morita and Hayashi, 1998; Morita, 1994). In contrast, dose-related increases in bone marrow micronuclei were reported by in male and female C57BL6 mice (Mirkova, 1994) at the same test conditions that gave negative results in this strain in Tinwell and Ashby (1994), and in CD1 mice (Roy et al., 2005).

All three studies of micronucleus formation in the liver, which is a target tissue for 1,4-dioxane carcinogenicity, were positive. Micronucleus formation in male CD1 mice was increased at doses  $\geq 2500$  mg/kg/day (Roy et al., 2005) and at  $> 2000$  mg/kg/day following partial hepatectomy to induce cellular mitosis (Morita and Hayashi, 1998). Roy et al. (2005) further investigated the origin of the micronuclei in bone marrow and liver. They concluded that 1,4-dioxane causes micronuclei formation primarily through chromosomal breakage, and that the compound can interfere with cell proliferation in both the liver and bone marrow. Itoh and Hattori (2019) recently reported that 1,4-dioxane increased hepatic micronuclei formation in male F344/DuCrIj rats after partial hepatectomy studies using three different methods (juvenile method using 4-week-old rats, dosing before or after partial hepatectomy using 8 week old rats). In contrast, 1,4-dioxane did not increase bone marrow micronuclei in this study, and the *Pig-a* assay, which detects inactivating mutations in an X-linked reporter gene in peripheral blood cells, was also negative.

Gi et al. (2018) evaluated mutagenicity and other endpoints related to the mechanism of hepatic carcinogenicity of 1,4-dioxane in *gpt* delta transgenic rats, a model for detection of *in vivo* mutations. Their overall conclusions from the studies, described below, were that “1,4-dioxane is a genotoxic hepatocarcinogen and induces hepatocarcinogenesis through a mutagenic mode of action rats.” Male *gpt* delta transgenic rats were given 0, 200, 1000, and 5000 ppm and wild type (WT) rats were given 0, 2, 20, 300, 2000, and 5000 ppm in their drinking water for 16 weeks, and daily doses in mg/kg/day were calculated from body weight and water consumption data. At the highest dose (5000 ppm - 440 mg/kg/day in transgenic rats, 562 mg/kg/day in WT rats), body weight was significantly decreased in WT and transgenic rats, and relative liver weight was slightly increased in transgenic rats. Histopathological changes including hypertrophy, swelling, necrosis, apoptosis or fatty changes were not seen in the liver at any dose in either strain. Glutathione S-transferase placental form (GST-P) foci, a preneoplastic lesion in rat liver, were increased at 2000 ppm (222 mg/kg/day) and 5000 ppm (562 mg/kg/day) in WT rats, and at 5000 ppm (440 mg/kg/day) but not at lower doses ( $\leq 92$  mg/kg/day) in transgenic rats. The BrdU labeling index, an indicator of proliferating cells, was significantly evaluated in livers of WT rats

at 5000 ppm, but not at lower doses; it was not evaluated in transgenic rats. The effect of 1,4-dioxane on mutation frequency and the types of mutations in the *gpt* gene was evaluated in the livers of transgenic mice. Mutation frequency, and A:T – G:C transition and A:T – T:A transversion mutations, were increased at 5000 ppm (440 mg/kg/day), and A:T – T:A transversion mutations were also increased at 1000 ppm (92 mg/kg/day). In gene expression studies in livers from transgenic rats, expression of genes involved in cell proliferation (proliferating cell nuclear antigen) and DNA damage repair (O-6-methylguanine-DNA methyltransferase) were increased at 5000 ppm (440 mg/kg/day). Based on the above, Gi et al. (2018) conclude that 1,4-dioxane is mutagenic to liver, its target organ in rats, and that the discrepancy between the negative *in vitro* mutagenicity studies and their positive *in vivo* study may arise from “organ-specific pathways of xenobiotic metabolism and DNA repair *in vivo*.”

Kitchin and Brown (1990) evaluated biochemical and histopathological changes in female Sprague-Dawley rats dosed with 1,4-dioxane by gavage. Single-strand DNA breaks in hepatocytes were increased at a dose that did not cause histopathological changes. At the doses that caused DNA damage in this study, ornithine decarboxylase and cytochrome P450 were also elevated. These two parameters were stated to be associated with tumor promotion, and the authors suggested that promotion may be involved with 1,4-dioxane carcinogenicity (Kitchin and Brown, 1990).

Hepatocyte DNA synthesis, indicative of cell proliferation, was also increased in several *in vivo* studies (Miyagawa et al., 1999; Uno et al., 1994; Goldsworthy et al., 1991; Stott et al., 1981). However, DNA repair in hepatocytes after *in vitro* or *in vivo* exposure and DNA repair in the nasal cavity after *in vivo* exposure were not affected by 1,4-dioxane (Goldsworthy et al., 1991; Stott et al., 1981), although, as discussed above, Gi et al. (2018) reported increased hepatic expression of a gene indicative of DNA repair.

Additional studies reported that 1,4-dioxane caused transient inhibition of RNA polymerase A and B in rat liver (Kurl et al., 1981), and that DNA alkylation was not detected in the liver of Sprague Dawley rats after gavage exposure (Stott et al., 1981).

Finally, a recent study (Furihata et al., 2018) compared effects of 1,4-dioxane, two genotoxic hepatic carcinogens (N-nitrosodiethylamine; 3,3'-dimethylbenzidine), and a non-genotoxic hepatic carcinogen (di[2-ethylhexyl]phthalate) on hepatic expression of 11 “marker genes” stated to discriminate genotoxic and non-genotoxic hepatic carcinogens. In this study, male F344 rats were dosed with 0.5% 1,4-dioxane in drinking water for 4 weeks. The gene expression profile of 1,4-dioxane was “intermediate” between that of the “typical” genotoxic and non-genotoxic to which it was compared.

In summary, while many of the genotoxicity studies reviewed above were negative, others provide evidence for mutagenicity and chromosomal damage, including some *in vivo* studies.

Notably, a recent in vivo study (Gi et al., 2018 – discussed below) provides evidence that 1,4-dioxane causes mutations in rat liver. Additionally, 1,4-dioxane induced micronuclei in the liver in all three studies where this effect was evaluated.

### **Tumor initiation and promotion studies**

Bull et al. (1986) reported that 1,4-dioxane was negative for cancer initiation in an initiator/promoter test in female SENCAR mice (6–8 weeks old). The mice were administered a single dose of 1,000 mg/kg by gavage, subcutaneous injection or topical application, followed by dermal application of a tumor promoter (1 µg of 12-O-tetradecanoylphorbol-13-acetate; TPA) or acetone (control) 3 times per week for 20 weeks. At 24 weeks, the formation of papillomas was not increased in mice treated with 1,4-dioxane and the promoter, and no tumors occurred in mice treated only with only 1,4-dioxane.

In a study in mice by King et al. (1973), 1,4-dioxane was negative as a complete carcinogen and was positive as a tumor promoter. Swiss Webster mice (30 per sex per treatment group) were dosed dermally with: 0.2 ml of a solution of 1,4-dioxane (concentration not provided) 3 times/week for 78 weeks; 0.2 ml of a solution of 1,4-dioxane 3 times/week for 78 weeks and with a tumor initiator (50 µg of dimethylbenzanthracene) one week before 1,4-dioxane dosing began; or only with the tumor initiator. In mice dosed with both the initiator and 1,4-dioxane, only 4 male and 5 female mice survived for 60 weeks (compared to 22 males and 25 females dosed only with 1,4-dioxane, and 20 males and 26 females dosed only with the initiator). 1,4-dioxane did not cause skin tumors in the absence of the initiator. In contrast, the incidence and multiplicity of skin tumors was higher in mice treated with both 1,4-dioxane and the initiator than in mice treated only with the initiator. Tumors of the lung and kidney also occurred in mice treated with 1,4-dioxane and the initiator.

Lundberg et al. (1987) reported that 1,4-dioxane is a promoter of liver tumors in rats. Partially hepatectomized male Sprague Dawley rats (9-11 per group) were dosed with: a tumor initiator (diethylnitrosamine, 30 mg/kg by intraperitoneal injection); 1,4-dioxane at 100 or 1,000 mg/kg/day by gavage, 5 days per week for 7 weeks; the initiator and 1,4-dioxane; or neither compound (controls). When evaluated 10 days after the last dose of 1,4-dioxane, the number and volume of hepatic GGT-foci were increased in rats dosed with both the initiator and 1,000 mg/kg/day 1,4-dioxane as compared to the rats dosed with only the initiator. Histopathological changes including enlarged, foamy hepatocytes containing numerous fat-containing cytoplasmic vacuoles were observed in the livers of rats dosed with 1,000 mg/kg/day 1,4-dioxane, regardless of whether or not they had been treated with the initiator.

Finally, as mentioned above, Kitchin and Brown (1990) reported that hepatic ornithine decarboxylase and cytochrome P450, which were stated to be associated with tumor promotion, were elevated in female Sprague-Dawley rats at a 1,4-dioxane dose that caused single-strand

DNA breaks but did not cause histopathological changes in hepatocytes (Kitchin and Brown, 1990).

## **REFERENCE DOSE FOR NON-CANCER EFFECTS AND CANCER SLOPE FACTOR**

### **USEPA (2013) Reference Dose for non-carcinogenic effects**

As mentioned above, USEPA (2018) and numerous states have used the USEPA (2013) cancer slope factor as the basis for their 1,4-dioxane drinking water guidelines, and the Health Effects Subcommittee agrees that the MCLG should be based on this approach. The USEPA (2013) Reference Dose for non-carcinogenic effects of oral exposure to 1,4-dioxane is presented here for the sake of completeness.

USEPA (2013) identified histopathological lesions of the liver and kidney (renal tubular epithelial and hepatocellular degeneration and necrosis) in rats in the Kociba et al. (1974) drinking water study as the most sensitive non-carcinogenic effect of oral exposure to 1,4-dioxane. Kociba et al. (1974) reported that these effects occurred in the mid- and high-dose groups but not in the control or low-dose groups, but they did not provide incidence data for the mid- and high-dose groups. The USEPA (2013) Reference Dose is based on the mean daily doses in males (0, 9.6, 94, and 1,015 mg/kg/day) reported by Kociba et al. (1974), since it is assumed these lesions occurred in both sexes and that the doses in males were lower than the doses in females (0, 19, 148, and 1,599 mg/kg/day).

Because the incidence of the lesions was not reported, Benchmark Dose (BMD) modeling could not be used in RfD development. Therefore, the NOAEL of 9.6 mg/kg/day was used as the point of departure (POD) for the Reference Dose. A total uncertainty factor of 300 (10 for interindividual variation; 10 for animal-to-human extrapolation; 3 for database deficiencies including lack of a multigeneration reproductive toxicity study) was applied to the NOAEL of 9.6 mg/kg/day to derive the RfD of 0.03 mg/kg/day.

### **Weight of evidence for carcinogenicity**

Data relevant to carcinogenicity of 1,4-dioxane (summarized above) come from seven studies in rats (five drinking water; two inhalation), two drinking water studies in mice, and one drinking water study in guinea pigs. Sites at which tumors were increased in at least one dosed group in these studies are shown in Table 3 above. In summary, liver tumors were increased in the studies of rats, mice and guinea pigs, with the exception of female rats in the NCI (1978) drinking water study and both sexes of rats in the Torkelson et al. (1974) inhalation study. Additionally, nasal cavity tumors in rats were increased in four drinking water studies and one inhalation study, and USEPA (2013) notes that these tumors were not evaluated in the other rat inhalation study (Torkelson et al., 1974). Increases of tumors at several other sites (mammary gland, mesothelioma of the peritoneum or testis/epididymis, kidney, zymbal gland, subcutis fibroma) were also reported in one or more rat studies.

1,4-Dioxane is described by USEPA (2013) as “likely to be carcinogenic to humans” as defined in the USEPA (2005) Guidelines for Carcinogen Risk Assessment. The Health Effects Subcommittee agrees with this USEPA (2013) conclusion, and it notes that the data shown in Table 3 clearly show that 1,4-dioxane fulfills the following USEPA (2005) criterion for this descriptor: “...tested positive in animal experiments in more than one species, sex, strain, site, or exposure route, with or without evidence of carcinogenicity in humans.” The Health Effects Subcommittee also agrees with USEPA (2013) that the “likely to be carcinogenic to humans” descriptor applies to all routes of exposure, since 1,4-dioxane caused tumors at sites remote from the portal of entry/site of absorption in oral and inhalation studies. Similarly, the International Agency for Research on Cancer (IARC, 1999) classified 1,4-dioxane as Group 2B (possibly carcinogenic to humans) based on inadequate evidence of carcinogenicity in humans and sufficient evidence of carcinogenicity in experimental animals, and the National Toxicology Program (NTP, 2011) concluded that 1,4-dioxane is “reasonably anticipated to be a human carcinogen based on sufficient evidence of carcinogenicity from studies of experimental animals.”

#### **Selection of dose-response approach for cancer risk assessment**

According to the USEPA (2005) Guidelines for Carcinogen Risk Assessment, an approach based on a non-threshold dose-response relationship (i.e. linear low-dose extrapolation; cancer slope factor) is used when the mode of action for carcinogenicity has not been conclusively established, or if a mutagenic mode of action has been established. An approach based on a threshold dose-response relationship (i.e. a Reference Dose) is used when a mode of action for carcinogenicity that is not linear at low doses (i.e. a threshold for carcinogenicity exists) has been clearly established. Information relevant to the choice of non-threshold or threshold approach for cancer risk assessment of 1,4-dioxane is discussed below.

USEPA IRIS (2013), USEPA Office of Chemical Safety and Pollution Prevention (OCSP; 2019), NJDEP (2015, 2018), and the other states listed in Table 1 all base their risk assessments for 1,4-dioxane on a non-threshold dose-response (i.e. a slope factor). In contrast, Health Canada (2018) concluded that “analysis supports a non-genotoxic mode of action involving cytotoxicity followed by regenerative hyperplasia and stimulation of endogenously formed mutations. Since 1,4-dioxane acts through a non-genotoxic mode of action and is known to operate via non-linear kinetics, a non-linear (threshold) risk assessment is considered appropriate.” It is noted that Health Canada’s decision relied heavily on the conclusions of Dourson et al. (2014, 2017), which are discussed in detail below.

The Health Effects Subcommittee reviewed two publications by Dourson et al. (2014, 2017) that conclude that 1,4-dioxane is carcinogenic through a threshold mode of action (MOA) and that the default non-threshold (linear low-dose extrapolation) approach for cancer risk assessment is therefore not appropriate. NJDEP (2015, 2018) responded to the conclusions of these two

publications when they were submitted in comments on the draft Interim Ground Water Quality Standard for 1,4-dioxane in 2015 (Dourson et al., 2014) and the proposed Ground Water Quality Standard for 1,4-dioxane in 2018 (Dourson et al., 2017). NJDEP (2015, 2018) concluded that the mode of action of 1,4-dioxane carcinogenicity remains unknown, and that the analyses presented in Dourson et al. (2014; 2017) do not conclusively establish a threshold mode of action. Similarly, USEPA (2019) also reviewed Dourson et al. (2014, 2017) and other relevant data and concluded that a threshold mode of action has not been established and that a non-threshold (slope factor) approach should be used for cancer risk assessment of 1,4-dioxane. The Health Effect Subcommittee agrees with NJDEP (2018) that “the data and explanations provided by Dourson et al. (2017) do not establish a firm or unique link to the proposed mode of action, and they do not indicate that a threshold approach is appropriate for risk assessment for [1,4-dioxane].” Health Effects Subcommittees conclusions about specific points presented by Dourson et al. (2014, 2017) are discussed in Appendix 1.

As shown in Table 3 above, 1,4-dioxane increased the incidence of nasal tumors in rats exposed orally and through inhalation, suggesting that these tumors do not occur only as a point of contact effect. Kasai et al. (2009) suggested a mode of action involving both induction of metabolic enzymes, cytotoxicity, and regenerative cell proliferation and genotoxicity. However, as noted by USEPA (2013), Kasai et al. (2009) did not observe cytotoxicity in the nasal cavity. USEPA (2013) further notes that “nasal lesions, including inflammation, hyperplasia, and metaplasia, were frequently seen in inhalation studies conducted by the NTP with no evidence of nasal carcinogenicity...” The Health Effects Subcommittee agrees with the USEPA (2013) conclusion that, based on the information discussed above, the mode of action for the nasal tumors has not been established. Similarly, the mode of action for other tumors reported to have been caused by 1,4-dioxane (kidney, lung, peritoneal mesothelioma, mammary gland, Zymbal gland, subcutis tumors) has not been established.

In summary, the mode of action for 1,4-dioxane carcinogenicity has not been conclusively established, and the USEPA (2005) Guidelines for Carcinogen Risk Assessment specify that a non-threshold approach (i.e. linear low-dose extrapolation; cancer slope factor) is to be used in such cases. Therefore, the Health Effects Subcommittee agrees with the USEPA (2103a) decision to use linear low-dose extrapolation (a cancer slope factor) for cancer risk assessment of 1,4-dioxane.

### **Cancer slope factor derivation**

Table 4 provides incidence data for liver tumors in mice and rats, and for nasal cavity, peritoneal, and mammary gland tumors in rats, from the Kociba et al. (1974), NCI (1978), and Kano et al. (2009) studies of exposure to 1,4-dioxane in drinking water.



**Table 4:** Incidence of liver, nasal cavity, peritoneal, and mammary gland tumors in rats and mice exposed to 1,4-dioxane in drinking water for 2 years (based on survival to 12 months) (USEPA, 2013)

Study	Species/strain/sex	Dose (mg/kg/day)	Tumor Incidence			
			Liver	Nasal Cavity	Peritoneal	Mammary gland
Kociba et al. (1974)	Sherman rats, male and female combined <sup>a,b</sup>	0	1/106 <sup>h</sup>	0/106 <sup>h</sup>	NA	NA
		14	0/110	0/110	NA	NA
		121	1/106	0/106	NA	NA
		1307	10/66 <sup>i</sup>	3/66	NA	NA
NCI (1978)	Male Osborne-Mendel rats <sup>b</sup>	0	NA	0/33 <sup>h</sup>	NA	NA
		240	NA	12/26	NA	NA
		530	NA	16/33 <sup>i</sup>	NA	NA
	Female Osborne-Mendel rats <sup>b,c</sup>	0	0/31 <sup>h</sup>	0/34 <sup>h</sup>	NA	NA
		350	10/30	10/30 <sup>i</sup>	NA	NA
		640	11/29	8/29 <sup>i</sup>	NA	NA
	Male B6C3F1 mice <sup>d</sup>	0	8/49 <sup>h</sup>	NA	NA	NA
		720	19/50 <sup>i</sup>	NA	NA	NA
		830	28/47 <sup>i</sup>	NA	NA	NA
	Female B6C3F1 mice <sup>d</sup>	0	0/50 <sup>h</sup>	NA	NA	NA
		380	21/48 <sup>i</sup>	NA	NA	NA
		860	35/37 <sup>i</sup>	NA	NA	NA
Kano et al. (2009)	Male F344/DuCrj rats <sup>d,e,f,g</sup>	0	3/50	0/50	1/50	8/50
		11	4/50	0/50	0/50	8/50
		55	7/50	0/50	0/50	11/50
		274	39/50 <sup>j,k</sup>	7/50 <sup>j,k</sup>	0/50	18/50 <sup>j,k</sup>
	Female F344/DuCrj rats <sup>d,e,f,g</sup>	0	23/50	0/50	NA	NA
		18	31/50	0/50	NA	NA
		83	37/50	0/50	NA	NA
		429	40/50	8/50	NA	NA
	Male Crj:BDF1 mice <sup>d</sup>	0	5/50	0/50	NA	NA
		49	35/50	0/50	NA	NA
		191	37/50 <sup>i</sup>	0/50	NA	NA
	Female Crj:BDF1 mice <sup>d</sup>	677	40/50 <sup>j,k</sup>	1/50	NA	NA
		0	5/50	0/50	NA	NA
		66	35/50 <sup>j</sup>	0/50	NA	NA
		278	41/50 <sup>j</sup>	0/50	NA	NA
	964	46/50 <sup>j,k</sup>	1/50	NA	NA	

<sup>a</sup> Incidence of hepatocellular carcinoma.

<sup>b</sup> Incidence of nasal squamous cell carcinoma.

<sup>c</sup> Incidence of hepatocellular adenoma.

<sup>d</sup> Incidence of hepatocellular adenoma or carcinoma.

<sup>e</sup> Incidence of all types of nasal tumors combined.

<sup>f</sup> Incidence of peritoneal mesotheliomas.

<sup>g</sup> Incidence of mammary gland fibroadenomas or carcinomas.

<sup>h</sup>  $p < 0.05$ ; positive dose-related trend (Cochran-Armitage or Peto's test).

<sup>i</sup> Significantly different from control at  $p < 0.05$  by Fisher's exact test.

<sup>j</sup> Significantly different from control at  $p < 0.01$  by Fisher's exact test.

<sup>k</sup>  $p < 0.01$ ; positive dose-related trend (Peto test).

NA = data not available for modeling.

Benchmark Dose (BMD) modeling was performed by USEPA (2013) using the dichotomous models included in Benchmark Dose Software (BMDS, version 2.1.1) for the data on the incidence of liver tumors (hepatocellular carcinoma or adenoma) in rats and mice, and nasal

tumors, peritoneal mesotheliomas, and mammary gland tumors in rats from Kano et al. (2009), NCI (1978), and Kociba et al. (1974). When deriving a cancer slope factor, the point of departure is the BMDL, which is the 95% lower confidence limit on the dose associated with a benchmark response (BMR) near the lower end of the observed data from the study. For 1,4-dioxane, modeling was performed using a BMR of 10%, and, as discussed below, additional modeling based on BMRs of 30% and 50% was conducted for the female mouse hepatic tumor data from Kano et al. (2009). The BMD (dose associated with the BMR) and BMDLs were first calculated based on the doses administered to the animals. The BMDs and BMDLs based on the animal doses were then converted by USEPA (2013) to the BMD<sub>HED</sub> and BMDL<sub>HED</sub>, which are the BMDs and BMDLs based on Human Equivalent Doses, using the default body weight (BW) scaling factor of BW<sup>0.75</sup> (U.S. EPA, 2011b), time-weighted average animal body weight data, and an assumed human body weight of 70 kg, as follows:

$$\text{HED} = \text{animal dose (mg/kg)} \times (\text{animal BW [kg]/human BW [kg]})^{0.25}$$

The dose-response data (Table 4-above) and cancer slope factors (Table 5-below) indicate that liver tumors in female mice (observed in both Kano et al., 2009 and NCI, 1978) are more sensitive to 1,4-dioxane than the other tumor types observed in rats and mice in Kano et al. (2009), NCI (1978) and Kociba et al. (1974). USEPA (2013) selected the hepatic tumors in female mice in the drinking water study conducted by Kano et al. (2009) as the basis for the cancer slope factor. NCI (1978) was not selected by USEPA (2013) because it included only two dose levels while Kano et al. (2009) used three dose levels, and because the lowest dose in NCI (1978) was much higher than in Kano et al. (2009). Kociba et al. (1974) was not selected by USEPA (2013) because it did not include mice and reported only hepatocellular carcinomas but not adenomas. In regard to the use of the Kano et al. (2009) mouse liver tumor data as the basis for risk assessment, USEPA (2013) notes that the background incidence of liver tumors is similar in the BDF1 strain of mice used by Kano et al. (2009) as in the B6C3F1 strain used by the National Toxicology Program and concludes that “the BDF1 mouse is not particularly sensitive compared to the commonly used B6C3F1 strain” (USEPA, 2013).

The dose-response curve for the female mouse hepatic tumor data from Kano et al. (2009) is very steep at the low dose and plateaus at a very high tumor incidences in the two higher doses; control, low-, mid-, and high-dose incidence are 10%, 70%, 82%, and 92% respectively. The log-logistic model was the only model that provided an adequate fit to these data (USEPA, 2013). Since the response level (70%) at the lowest dose in the study (Kano et al., 2009) was much higher than the initial BMR of 10%, modeling was also performed using the log-logistic model for BMRs of 30 and 50%. USEPA (2013) selected the human equivalent dose BMDL for a BMR of 50% (BMDL<sub>50-HED</sub> of 4.95 mg/kg/day) for female mouse liver tumors in Kano et al., 2009 as the point of departure for deriving for the cancer slope factor. The slope factor was calculated as follows:

$$\text{CSF} = \text{BMR/BMDL}_{50\text{-HED}} = 0.5 / 4.95 \text{ mg/kg/day} = 0.10 \text{ (mg/kg/day)}^{-1}$$

Table 5. Oral cancer slope factors for best-fit models for tumor incidence data for rats and mice exposed to 1,4-dioxane in drinking water for 2 years (*adapted from USEPA, 2013*)

<i>Study</i>	<i>Gender/strain/species</i>	<i>Tumor type</i>	<i>BMR</i>	<i>Oral Cancer Slope Factor (mg/kg/day)<sup>-1</sup></i>	<i>Model</i>
Kano et al. (2009)	Male F344/DuCrj rats	Hepatocellular adenoma or carcinoma	0.1	0.007	Probit, slope parameter not restricted
	Female F344/DuCrj rats		0.1	0.0069	Multistage, degree of polynomial=2
	Male Crj:BDF1 mice		0.1	0.037	Log-logistic, slope restricted $\geq 1$
	Female Crj:BDF1 mice		0.1	0.18	
			0.3	0.14	
			0.5	0.1	
	Female F344/DuCrj rats	Nasal squamous cell carcinoma	0.1	0.0014	Multistage, degree of polynomial=3
	Male F344/DuCrj rats		0.1	0.0015	
Male F344/DuCrj rats	Peritoneal mesothelioma	0.1	0.0047	Probit, slope parameter not restricted	
Female F344/DuCrj rats	Mammary gland adenoma	0.1	0.0049	Log-logistic, slope restricted $\geq 1$	
Kociba et al. (1974)	Male and female combined Sherman rats	Nasal squamous cell carcinoma	0.1	0.00029	Multistage, degree of polynomial=3
		Hepatocellular carcinoma	0.1	0.00042	Probit, slope parameter not restricted
NCI (1978)	Male Osborne-Mendel rats	Nasal squamous cell carcinoma	0.1	0.0094	Log-logistic, slope restricted $\geq 1$
	Female Osborne-Mendel rats		0.1	0.0039	
	Female Osborne-Mendel rats	Hepatocellular adenoma	0.1	0.0054	
	Female B6C3F1 mice	Hepatocellular adenoma or carcinoma	0.1	0.01	Multistage, degree of polynomial=2
	Male B6C3F1 mice		0.1	0.0028	Gamma

## **DEVELOPMENT OF RECOMMENDED HEALTH-BASED MAXIMUM CONTAMINANT LEVEL**

### **Updated drinking water exposure assumptions**

The USEPA (2015) has updated its default assumptions for body weight and drinking water consumption used in calculation of health-based water values. USEPA (2015) updated the default adult body weight from 70 kg to 80 kg based on the mean body weight for adults age 21 and older in National Health and Nutrition Examination Survey (NHANES) from 1999-2006 (USEPA, 2011). The previous value of 70 kg was stated by USEPA (2015) to have been based on the mean adult body weight from NHANES III (1988-1994). USEPA (2015) also updated the default drinking water consumption rate to 2.4 L/day based on the estimated 90<sup>th</sup> percentile of community water ingestion for adults ages 21 and older in NHANES 2003-2006 (USEPA, 2011b). The previous value of 2 L/day was stated by USEPA (2015) to have been based on the 86<sup>th</sup> percentile of community water ingestion for adults from the US Department of Agriculture's 1994-1996 Continuing Survey of Food Intake by Individuals (CSFII) analysis and the 88<sup>th</sup> percentile of adults in the National Cancer Institute study of the 1977-1978 Nationwide Food Consumption Survey. These updated values were used by the Health Effects Subcommittee to develop the Health-based MCL for 1,4-dioxane, and they will also be used when Health-based MCLs for other contaminants are developed in the future.

### **Health-based MCL based on non-cancer effects**

As discussed above, it is well established that non-carcinogenic effects are less sensitive endpoints than carcinogenicity for 1,4-dioxane. The health-based MCL based on the Reference Dose for non-cancer effects (0.03 mg/kg/day; 30 µg/kg/day) and default exposure assumptions is presented here for comparison purposes.

$$\frac{30 \mu\text{g/kg/day} \times 80 \text{ kg} \times 0.2}{2.4 \text{ L/day}} = 200 \mu\text{g/L}$$

Where: 80 kg is the assumed body weight of an adult, 2.4 L/day is the default value for daily water consumption of an adult, and 0.2 (20%) is the default Relative Source Contribution factor.

### **Health-based MCL based on carcinogenicity**

The Health-based MCL for 1,4-dioxane is based on the one-in-one million (10<sup>-6</sup>) risk of cancer from lifetime exposure to carcinogens specified in the 1984 Amendments to the New Jersey Safe Drinking Water Act (SDWA) at N.J.S.A. 58:12A-20.

The daily dose of 1,4-dioxane predicted to result in a one-in-one-million lifetime cancer risk is calculated from the slope factor of 0.10 (mg/kg/day)<sup>-1</sup> as:

$$10^{-6} / 0.1 \text{ (mg/kg/day)}^{-1} = 1 \times 10^{-5} \text{ mg/kg/day} = 0.01 \mu\text{g/kg/day}$$

The Health-based Maximum Contaminant Level for 1,4-dioxane based on this daily dose is:

$$\frac{0.01 \mu\text{g}/\text{kg}/\text{day} \times 80 \text{ kg}}{2.4 \text{ L}/\text{day}} = 0.33 \mu\text{g}/\text{L}$$

Where: 80 kg is the assumed body weight of an adult and 2.4 L/day is the default value for daily water consumption of an adult.

This Health-based MCL is far below the Health-based MCL based on non-carcinogenic effects of 200  $\mu\text{g}/\text{L}$ .

**RECOMMENDATION:**

The recommended Health-based Maximum Contaminant Level for 1,4-dioxane is 0.33  $\mu\text{g}/\text{L}$ .

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**APPENDIX 1: REVIEW OF DOURSON ET AL. (2014) AND DOURSON ET AL. (2017)**

The Health Effects Subcommittee reviewed two publications by Dourson et al. (2014, 2017) that conclude that 1,4-dioxane is carcinogenic through a threshold mode of action (MOA) and that the default non-threshold (linear low-dose extrapolation) approach for cancer risk assessment is therefore not appropriate. NJDEP (2015, 2018) responded to the conclusions of these two publications when they were submitted in comments on the draft Interim Ground Water Quality Standard for 1,4-dioxane in 2015 (Dourson et al., 2014) and the proposed Ground Water Quality Standard for 1,4-dioxane in 2018 (Dourson et al., 2017). NJDEP (2015, 2018) concluded that the mode of action of 1,4-dioxane carcinogenicity remains unknown, and that the analyses presented in Dourson et al. (2014; 2017) do not conclusively establish a threshold mode of action. Similarly, USEPA (2019) also reviewed Dourson et al. (2014, 2017) and other relevant data and concluded that a threshold mode of action has not been established and that a non-threshold (slope factor) approach should be used for cancer risk assessment of 1,4-dioxane.

Health Effects Subcommittee conclusions about specific points made by Dourson et al. (2014, 2017) are presented below:

- **Dourson et al. (2014, 2017)** conclude that 1,4-dioxane causes tumors through a threshold mode of action involving cytotoxicity, necrosis, and regenerative hyperplasia followed by tumor formation.

**The Health Effects Subcommittee** finds that the Dourson et al. (2014, 2017) analyses do not conclusively establish a threshold mode of action for 1,4-dioxane carcinogenicity. The Subcommittee's conclusions are consistent with those of NJDEP (2015b, 2018) and USEPA (2019). Relevant to this issue, as summarized in Table 4, liver tumors in several rodent studies occurred at doses at which there were no lesions indicative of cytotoxicity and/or regeneration (Kano et al., 2009; NCI, 1978). These data indicate that 1,4-dioxane can cause liver tumors in the absence of cytotoxicity followed by cell proliferation.

<b>Table 4. Temporal sequence and dose-response relationship for possible key events and liver tumors in rats and mice. Adapted from USEPA, 2013.</b>				
<b>Dose (mg/kg-day) or Exposure (ppm)</b>	<b>Key event (time →)</b>			
	<b>Liver damage</b>	<b>Cell proliferation</b>	<b>Hyperplasia</b>	<b>Adenomas and/or carcinomas</b>
<b>Kociba et al., (1974)—Sherman rats (male and female combined)</b>				
0 mg/kg-day	__a	__a	__a	__a
14 mg/kg-day	__a	__a	__a	__a
121 mg/kg-day	+ <sup>c</sup>	__a	+ <sup>c</sup>	__a
1,307 mg/kg-day	+ <sup>c</sup>	__a	+ <sup>c</sup>	+ <sup>c</sup>
<b>NCI, (1978)—female Osborne-Mendel rats</b>				
0 mg/kg-day	__a	__a	__a	__a

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350 mg/kg-day	__a	__a	__a	+ <sup>c</sup>
640 mg/kg-day	__a	__a	__a	+ <sup>c</sup>
<b>NCI, (1978)—female Osborne-Mendel rats</b>				
0 mg/kg-day	__a	__a	__a	__a
720 mg/kg-day	__a	__a	__a	+ <sup>c</sup>
830 mg/kg-day	__a	__a	__a	+ <sup>c</sup>
<b>NCI, (1978)—male B6C3F<sub>1</sub> mice</b>				
0 mg/kg-day	__a	__a	__a	__a
720 mg/kg-day	__a	__a	__a	+ <sup>c</sup>
830 mg/kg-day	__a	__a	__a	+ <sup>c</sup>
<b>NCI, (1978)—female B6C3F<sub>1</sub> mice</b>				
0 mg/kg-day	__a	__a	__a	__a
380 mg/kg-day	__a	__a	__a	+ <sup>c</sup>
860 mg/kg-day	__a	__a	__a	+ <sup>c</sup>
<b>Kano et al., (2009) —male F344/DuCrj rats</b>				
0 mg/kg-day	__a	__a	__a	__a
11 mg/kg-day	__a	__a	__a	__a
55 mg/kg-day	__a	__a	__a	__a
274 mg/kg-day	+ <sup>c,d</sup>	__a	__a	+ <sup>c,e</sup>
<b>Kano et al., (2009)—female F344/DuCrj rats</b>				
0 mg/kg-day	__a	__a	__a	__a
18 mg/kg-day	__a	__a	__a	__a
83 mg/kg-day	__a	__a	__a	__a
429 mg/kg-day	__a	__a	__a	+ <sup>c,e</sup>
<b>Kano et al., (2009)—male Crj:BDF1 mice</b>				
0 mg/kg-day	__a	__a	__a	__a
49 mg/kg-day	__a	__a	__a	+ <sup>c,e</sup>
191 mg/kg-day	__a	__a	__a	+ <sup>c,e</sup>
677 mg/kg-day	+ <sup>c,d</sup>	__a	__a	+ <sup>c,e</sup>
<b>Kano et al., (2009)—female Crj:BDF1 mice</b>				
0 mg/kg-day	__a	__a	__a	__a
66 mg/kg-day	__a	__a	__a	+ <sup>c,e</sup>
278 mg/kg-day	__a	__a	__a	+ <sup>c,e</sup>
964 mg/kg-day	+ <sup>c,d</sup>	__a	__a	+ <sup>c,e</sup>
<b>Kasai et al., (2009)—male F344 rats</b>				
0 ppm	__a	__a	__a	__a
50 ppm	__a	__a	__a	__a
250 ppm	__a	__a	__a	__a
1,250 ppm	+ <sup>h</sup>	__a	__a	+ <sup>h</sup>
<sup>a</sup> — No evidence demonstrating key event. <sup>b</sup> + 1,4-dioxane metabolism was not evaluated as part of the chronic bioassays. Data from pharmacokinetic studies suggest that metabolism of 1,4-dioxane by CYP2E1 and CYP2B2 occurs immediately and continues throughout the duration of exposure at all				

exposure levels.

<sup>c</sup>+ Statistically significant increase noted.

<sup>d</sup>+ Single cell necrosis was observed in a 13 week bioassay for male rats (274 mg/kg-day), male mice (585 mg/kg-day), and female mice (898 mg/kg-day) exposed to 1,4-dioxane in drinking water ([Kano et al., 2008](#)).

<sup>e</sup>+ Kano et al. ([2009](#)) reported incidence rates for hepatocellular adenomas and carcinomas.

<sup>f</sup>+ Kasai et al. ([2008](#)) reported significant incidence rates for single cell necrosis in female rats only (3,200 ppm) following a 2 year bioassay.

<sup>g</sup>— All rats died during the first week of the 13-week bioassay ([Kasai et al., 2008](#)).

<sup>h</sup>+ Kasai et al. ([2009](#)) reported incidence rates for centrilobular necrosis and hepatocellular adenomas in male rats (1,250 ppm).

- **Dourson et al. (2014)** conducted a pathology review of the mouse liver slides from the chronic oral study conducted by NCI (1978). In NCI (1978), there was a dose-related increase in the incidence of liver tumors in both male and female mice, as follows:

Males: Control-16%, 720 mg/kg/day-38%, 830 mg/kg/day-60%.

Females: Control-0%, 380 mg/kg/day- 44%, 860 mg/kg/day-95%.

Dourson et al. (2014) state that, at the time that the NCI (1978) study was conducted, only the most severe response seen on a slide was recorded, so that if a tumor was observed, non-neoplastic changes on the same slide would not have been noted. They suggest that non-neoplastic changes such as glycogen depletion, hypertrophy, necrosis, inflammation, and Kupffer cell hyperplasia preceded and were causative to tumor formation.

**The Health Effects Subcommittee** notes that, in the Dourson et al. (2014) pathology review, a higher incidence and/or greater severity for these non-neoplastic effects were observed in both the high and low dose male mice than in controls. However, the incidence and/or severity of the non-neoplastic changes in female mice was similar or greater in controls than in the low dose group, while the incidence of liver tumors in the control and low dose female mice were 0 and 44% respectively. Therefore, the Health Effects Subcommittee agrees with the NJDEP (2015) conclusion that these data suggest that such non-neoplastic changes are not part of the sequence of events leading to tumor formation in the low dose female mice.

- **Dourson et al. (2014)** that the incidence of non-neoplastic effects does not correlate with the tumor incidence in control and low-dose females in NCI (1978). That state that the lower incidence of non-neoplastic changes in low-dose females than in low-dose males may be due to the fact that the low dose in females was about half of the low dose in males.

**The Health Effects Subcommittee** agrees with the NJDEP (2015) conclusion that Dourson et al. (2014) do not provide a logical explanation since the incidence of liver tumors in low

dose females (44%, as compared to 0% in controls) is greater than in low dose males (38%, as compared to 16% in controls) at a dose almost 2-fold higher.

The Health Effects Subcommittee as notes that , as discussed in more detail below, the USEPA (2013) oral slope factor is based on female mouse liver tumors from Kano et al. (2009), not data from NCI (1978) study. In this study, a different strain of mice (Crj:BDF1) were used. In Kano et al. (2009), the low dose in female mice (66 mg/kg/day) was almost 6-fold lower than the low dose in NCI (1978) (380 mg/kg/day), and non-neoplastic changes such as necrosis were not observed in the liver. However, the tumor incidence in the low dose females (70% compared to 10% in controls) in Kano et al. (2009) was higher than at the much higher dose (44% at 380 mg/kg/day compared to 0% in controls) in the low dose females in NCI (1978). These data further support the conclusion that non-neoplastic changes do not necessarily precede hepatic tumors caused by 1,4-dioxane.

- **Dourson et al. (2014)** state that the incidence of non-neoplastic changes in the female controls may have been due to a viral infection that “was known to occur in all mice at the time of the bioassay.” They attribute this statement to E.E. McConnell, who conducted the pathology review and is a co-author of Dourson et al. (2014).

**The Health Effects Subcommittee** notes that no citation about the presence of the viral infection is provided, and this issue is not mentioned in either NCI (1978) or the pathology review report by Dr. McConnell. Additionally, this explanation is not logical, since it was not stated that the control females were specifically infected with the virus, as compared with other groups of male and female mice included in the study. The Health Effects Subcommittee agrees with the NJDEP (2015) conclusion that, if the pathway hypothesized by Dourson et al. (2014), in fact, resulted in tumors, then the presence of the elements of this pathway in control females, with incidence of necrosis and inflammation greater than in the low dose group, would also have been expected, regardless of its etiology, to result in tumors. The absence of tumors in the control females is thus inconsistent with the hypothesized link between the non-neoplastic changes observed in both control and treated mice and the observed tumors.

- **Dourson et al. (2014)** made the general conclusion, based on the points above, that non-neoplastic changes occur both more frequently at higher doses and are necessary precursors to tumor formation.

**The Health Effects Subcommittee** agrees with the NJDEP (2015) conclusion that, when considered as a whole, the information presented by Dourson et al. (2014) does not support the conclusions that non-neoplastic changes occur both more frequently at higher doses and are necessary precursors to tumor formation. In general, the data and explanation provided by Dourson et al. (2014) do not establish a firm or unique link to the proposed MOA of cytotoxicity followed by regenerative hyperplasia, and does not indicate that a threshold

approach is appropriate for risk assessment for this compound. As such, the information provided by Dourson et al. (2014) does not invalidate the conclusion made by USEPA IRIS (2013) that the available information does not establish a plausible mode of action for 1,4-dioxane, and that the available data are not sufficient to establish significant biological support for a non-linear (threshold) mode of action. For these reasons, the approach used by USEPA IRIS (2013) which uses a linear low dose extrapolation to develop an oral cancer slope factor for 1,4-dioxane is appropriate.

- **Dourson et al. (2017)** revisited the results of the Kano et al. (2008, 2009) 13 week and two-year drinking water studies in rats and mice and reviewed English translations of the original Japanese laboratory reports of these studies (JBRC, 1990a, b). Some of the analyses presented by Dourson et al. (2017) are based on pooled data from male and female rats and mice (i.e. dose-response analyses are based on data points from male groups combined with data points from female groups). Furthermore, Dourson et al. (2017) adjusted doses from the 13-week studies, dividing them by a factor of 3, for comparison with the doses at which effects occurred in the two-year study. The rationale for this adjustment was that when point of departure (NOAEL, LOAEL, BMDL) from a subchronic study is used as the basis for a chronic Reference Dose, an uncertainty factor of 3 or 10 is applied to account for potential effects at lower doses with longer exposure durations.

**The Health Effects Subcommittee** agrees with the NJDEP (2018) conclusion that this adjustment is not appropriate for quantitative comparison of dose-response relationships from studies of different durations as was done by Dourson et al. (2017). Therefore, conclusions from Dourson et al. (2017) based on such comparisons (e.g. that centrilobular swelling and single cell liver necrosis in the 13-week rat study occurred at lower doses than tumors in the two-year rat study) are not scientifically supportable. Additionally, the Subcommittee notes that, as acknowledged by Dourson et al. (2017), even with the subchronic dose adjusted downward by a factor of 3, liver tumors occurred in mice in the chronic studies at doses below those that caused liver swelling and necrosis in the subchronic studies.

- **Dourson et al. (2017)** also conclude that the toxicity pathway for 1,4-dioxane is dependent on saturation kinetics, with decreased metabolism of the parent compound leading to increased toxicity.

**The Health Effects Subcommittee** concluded that, in the chronic mouse study, liver tumors were increased in females at doses that did not cause the postulated “key events” including saturation of metabolism, as well as, increased liver weight/hypertrophy and necrosis/inflammation, that are required for the proposed threshold mode of action.

In summary, the Health Effect Subcommittee agrees with NJDEP (2018) that “the data and explanations provided by Dourson et al. (2017) do not establish a firm or unique link to the

proposed mode of action, and they do not indicate that a threshold approach is appropriate for risk assessment for [1,4-dioxane].”